

Multidrug Resistance and Plasmid Profile of *Staphylococcus Aureus* and *Escherichia Coli* Isolated From Patients Attending Urology Clinic, Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria

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Abstract

Urinary tract infections (UTIs) remain the most common infections that affect all age groups worldwide. This study was aimed at determining the multidrug resistance (MDR) and plasmid profile of *Staphylococcus aureus* and *Escherichia coli* among patients attending Urology Clinic of Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria. A total of two hundred and two (202) urine samples were collected in sterile containers and analyzed using standard microbiological techniques. Antibiotic susceptibility testing of the isolates was carried out using standard laboratory guidelines. Of the 202 samples analysed, 104 (51.4%) had significant bacteriuria. *Staphylococcus aureus* showed high resistance to Ampicillin, Amoxicillin, and Norfloxacin while *E. coli* was highly resistant to Rifampicin, Ampicillin, Ciprofloxacin and Ofloxacin. A total of 19 (18.27%) isolates were resistant to more than two different classes of antibiotics i.e. MDR. Out of the 19 MDR isolates, 11.29% were *S. aureus* while 19.35% were *E. coli*. These isolates were carrying plasmid for MDR ranging from 2500 base pair (bp) to 6000bp for *S. aureus* while 11,849bp for *E. coli*. Plasmids were therefore implicated to be responsible for drug resistance in most of the UTIs. Therefore, proper hygiene by individuals and public awareness policies by government agencies should be encouraged; and antibiotics for the treatment of UTIs should be based on these current antibiogram results.

Keywords: Urinary Tract Infections, Multidrug Resistance, Plasmid Profile, *Staphylococcus aureus*, *Escherichia coli*.

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INTRODUCTION

There are reports of illness and death related to increased rates of urinary tract infections caused by uropathogens and this results in increased rates of most hospital visits and related nosocomial infection of the beings [1,2]. There could be numerous causes of disease in beings, but infectious diseases are set whenever a pathogenic microbe is able to infiltrate host tissue, settle and cause harm to the tissues. The urinary tract or urinary system is made up of those structures or organs related with the production, transport, storage and discharge (voiding) of urine and infestation of any of its part by a pathogenic microorganism is called Urinary Tract Infection (UTI). These structures include: kidney, ureter, bladder, urethra, nerve cells and other structures [3,4].

Some of the clinical signs or symptoms of UTI includes burning feelings during urination (micturation), frequency in micturation or incontinence, loin back or abdominal pains, turbid or dark or

sometimes bloody urine, or even an unusual-smelling of urine accompanied by feverish condition. It is also possible to have nausea or vomiting sometimes when chronic stage is attained as reported by the National Kidney and Urologic Diseases Information Clearinghouse (NKUDIC) [5]. These pathogenic microbes could be bacteria, fungi, virus or even parasites [6]. Bacteria have been documented as the main uropathogen affecting the urinary tract [7].

An estimated people of more than 100 million world-wide came down with UTI annually [8]. This is perceived to be a burden to the nations and persons affected [9]. An approximated 1 billion US dollars is spent annually on cases of more than 8 million hospitalizations from emergency, hospital clinics and office visits in the United States [10]. The epidemiology of pathogenic UTI cases is considerably changing, claiming lives of persons owed to its development of resistance [11,12]. Virulence issues for UTI caused by *S. aureus* and *E. coli* include the specific surface

associated adhesions they possess, toxins they produce, polysaccharides contain on their cell wall, locomotory organelles and siderophores [13,14].

Predisposing factors to UTIs include: sexual intercourse with infected person [15], gender [16], urinary catheters [17, 18], inheritance, disease condition such as diabetes [15], uncircumcised persons [19], enlarged prostates [20], spinal cord injury due to use of catheter for voiding during urethral dysfunction [21]. Preceding exposure to UTI is also known to be a predisposing factor [22]. Underprivileged personal hygiene and cultural habits are also a predisposing factor for UTI [23].

The diagnosis of UTI is based on urinalysis, dipstick leukocyte esterase test for detecting blood in urine (10 WBC/mm^3); and the presence of Nitrite in a urine sample is an indication of UTI [24]. Nevertheless, urine microscopy, culture and sensitivity are the widely used and most preferred methods for diagnosis.

There is a global interest with regards to antimicrobial resistance in pathogenic organisms, with particular emphasis on *S. aureus* and *E. coli* [25]. There is the need to intensify efforts aimed at knowing the causative agents of UTIs in relation to their antimicrobial activities to the readily available drugs as this will help in the appropriate prescription policies, choice and use of antimicrobial agents within a locality [26]. The societies are now uninterruptedly faced with multidrug resistant species of organisms causing UTI [27]. This has been well-known to be a major problem and a major risk to health globally [28]. The multidrug resistant challenge has also been documented in Nigeria [29]. The unprincipled use and management of antimicrobial agents among patients and healthcare workers have led to an increase in microbial resistance among uropathogens with a resulting spread to other antibiotic susceptible organisms causing a serious health impediment [30, 31, 32]. The age, health status of an individual and the resistant nature of the species determines the nature of the infection [33]. Several cases of ill-health and renal abnormality or failure have been attributed to *S. aureus* and *E. coli* infections among urology patients [34].

Determination of plasmid is very imperative in epidemiological studies. Besides, it has been proved to be the earliest DNA-based method applied for analyzing resistance pattern, frequency and probable future status of the resistance in relation to certain parameter [35]. Molecular identification of genes offers the feature information about the pathways followed for resistance, effectiveness of bacteria to drive these possessions in long run and genotypic view of resistance whereas curing of plasmid and transfer explains frequency of resistance transferred to the extent of emerging resistance in novel strains [35].

Nevertheless, there are limited study on the multidrug resistance and plasmid profile of *S. aureus* and *E. coli* from UTI cases in this study area were known to authors, although, there was report on its prevalence in this study area [12]. Thus, this informed the choice of research area and the health facility. Therefore, this research work was aimed to examine the multidrug resistance and plasmid profile of *Staphylococcus aureus* and *Escherichia coli* isolated from patients attending Urology clinic of Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria. At the end of this study the bacteriuria status of the patients will be ascertained, susceptibility of the isolates from the urine samples of the patients will also be known and this will give rise to determine the plasmid profile embedded in those multidrug resistant organisms among the isolates.

MATERIALS AND METHODS

Study Area

The study was carried out in Lafia. Lafia is situated in the North central part of Nigeria, lying at latitude $8^{\circ}29'30''$. It has a total of 330, 712 inhabitants according to [36,37]. In another report by World Population Review (WPR), [38], Lafia has a population of 127, 236.

Ethical Consideration and approval

Ethical approval for this study was obtained from Dalhatu Araf Specialist Hospital ethical committee with protocol approved number DASH/L/ADM/0340.

Consent

The consent of each patient was sought and consent forms were signed to serve as an agreement of participation in the research work.

Collection of Samples

After duly informing the patients on how to collect their urine samples into sterile containers, two hundred and two (202) mid-stream urine specimens were collected from patients attending Urology Clinic of the hospital. The recruitment of the research patients was randomized. Samples were collected from patients between 6a.m. and 9a.m. into sterile bottles. The sample container caps were tightly closed then stored in ice packed containers for onward transport to the laboratory. To evade cross contamination during sampling, sterile gloves and forceps were used. The samples were immediately transported to the Microbiology laboratory of Federal of University, Lafia, for microbiological analysis.

Isolation and Identification of Test Organisms

The urine from the sterile sample containers were aseptically streaked onto Mannitol Salt Agar (MSA) and Eosin Methylene Blue Agar (EMBA) plates and incubated at 37°C for overnight as described by Cheesbrough, [39]. Greenish metallic sheen colonies

that grew on EMBA were carefully chosen as *E. coli* while those appearing yellowish with color change on MSA plates to pink color indicating fermentation of sugar were carefully chosen as *S. aureus*. The probable isolates were identified through microscopy (Gram stain) and biochemical tests particularly catalase, coagulase, indole and citrate tests respectively. The isolates were then stored on a nutrient agar slant for subsequent analysis.

Antibiotic Susceptibility Testing

The antibiotic susceptibility tests of *S. aureus* and *E. coli* isolates were carried out as done by Kirby Bauer disc diffusion method and modified by Helen and Jenifer, [40]. Briefly, 3 pure colonies of the isolated species from the patients' urine samples were inoculated into 5 ml sterile normal saline (0.85% w/v NaCl) and the turbidity of the bacteria solution was adjusted to equivalent that of 0.5 McFarland's standard solution (0.5ml of 1.172% w/v BaCl₂. 2H₂O into 99.5ml of 1% w/v H₂SO₄). Sterile swab sticks soaked in the standardized bacterial suspension were streaked onto already prepared Mueller-Hinton media and the antibiotic disks were then aseptically embedded onto the center of the inoculated Mueller-Hinton media. The effectiveness of the antibiotics against the test organisms were checked for either resistance, intermediate resistance or susceptible after comparing the inhibition zone diameter of each antibiotic by its corresponding clinical zone diameter break point (mm). The antibiotics tested against the isolates of *S. aureus* and *E. coli* were: Ofloxacin (10 µg), Levofloxacin (10 µg), Nalidixic acid (30 µg), Ciprofloxacin (10 µg), ampicillin/clavulanic acid (30 µg), Gentamycin (10 µg), Cotrimoxazole (30 µg), Streptomycin (30 µg), Cefalexin (10 µg), Ampicillin (30 µg), Ciprofloxacin (10 µg), Norfloxacin (10µg), Gentamycin (10µg), Streptomycin (30µg), Amoxicillin (20 µg), Erythromycin (30 µg), Rifampicin (20 µg), Chloramphenicol (30 µg), Ampicillin (20 µg), and Levofloxacin (20 µg) as described by Godwin *et al.* [41]. *Staphylococcus aureus* (NCTC 6571) and *Escherichia coli* (NCTC 10418) obtained from Department of Microbiology and Biotechnology, National Institute for Pharmaceutical Research and Development (NIPRD) were used as control.

Plasmid DNA Analyses of Isolates

Plasmid DNA analyses of the multi-drug resistant isolates were performed at Applied Biotechnology International Nigeria Limited (ABINL) Abuja, Nigeria.

Extraction of Plasmid DNA Using Spin-Column Protocol

Plasmid DNA extractions were done as described by Bimboim and Doly, [42] and Akinjogunla and Enabulele, [43] using Bacterial plasmid DNA miniprep™ (Zymo Scientific, USA). Each isolate was inoculated into 10 ml Luria-Bertani (LB) broth and

incubated for 16 hours at 37°C while shaking at 200-250rpm. The bacterial culture was harvested by centrifugation at 8000rpm in a microcentrifuge for two minutes at room temperature. The supernatant was decanted and all remaining medium removed. The pelleted cells were re-suspended in 250 µL of re-suspension solution and transferred to microcentrifuge tube for another round of centrifugation.

Exactly 250 µL of lysis solution was added and mixed thoroughly by inverting the tube 4-6 times until solution was viscous and slightly clear. This was followed by adding 350 µL of neutralization solution and mixed by inverting the tube. The samples were centrifuged at 10,000rpm for five minutes to pellet cell debris and chromosomal DNA. The supernatant was transferred to Gene JET spin column by decantation.

The samples were centrifuged within a minute and the supernatant were discarded. A solution of 500 µL was added to the column and centrifuged within a minute, flow-through were discarded and the column placed back into collection tube. The wash procedure was repeated to avoid residual ethanol in plasmid preps. The Gene JET spin column was transferred into fresh 1.5 mL microcentrifuge tube and 50µl of elution buffer was added to the centre of the column to elute plasmid DNA. This was incubated for 2 minutes at room temperature and centrifuged for two minutes. The cleansed plasmid DNA was stored at -20°C for further study.

Plasmid Curing

The isolates that were multiple drug resistant due to plasmid bands were exposed to plasmid curing using the method described by Akinjogunla and Enabulele, [43]. One (0.1) mg/mL of ethidium bromide was added into 100 mL of Luria Bertaru broth. The resolution was autoclaved at 121°C at 15 psi for 15 min. An overnight culture of the sample was standardized with 0.5 McFarland standard and 0.5 mL of the standardized suspension was pipette using pasteur pipette into 100 mL sterile Luria Bertaru broth. The solution was incubated at 37°C for 4 hrs. After the incubation period, the isolates were re-inoculated into a sterile nutrient broth and incubated for 24 hrs.

Quantification of the Extracted plasmid DNA

The extracted plasmids DNA were quantified using the Nanodrop 1,000 spectrophotometer. This was done by placing a drop i.e. approximately 2 µL on the sample space and analysed using the Nanodrop 1,000 software [9].

Detection of Number and Sizes of Plasmid DNA (Agarose Gel Electrophoresis)

One percent (1.0%) agarose gel was used to detect the plasmid DNA portions. This was prepared by combining 1g agarose in ten times concentration of tris-borate ethylenediaminetetraacetic acid (10 mL 10XTB-

EDTA) buffer and 90ml sterile distilled water in 250 mL beaker flask and heating in a microwave for 2 minutes until the agarose was dissolved [44].

Exactly, 0.5µL of Ethidium bromide was added to the dissolved agarose solution with swirling to mix. The gel was then poured onto a mini horizontal gel electrophoresis tank and casting combs were introduced. This was allowed to gel for thirty minutes. The casting combs were cautiously removed after the gel had solidified completely. One times concentration (1X) Tris Borate EDTA (TBE) buffer was added to the reservoir until it covered the agarose gel. Precisely 0.5 µL of gel tracking dye (bromophenol blue) was added to 20 µL of each sample with gentle mixing. Twenty microliters volume of the sample was loaded onto the wells of the gel, the mini horizontal electrophoresis gel setup was covered and electrodes connected. Electrophoresis was performed at 100 – 200mA for one hour. At the completion of electrophoresis, the gel was removed from the buffer and viewed under UV-transilluminator. The band pattern of DNA portions was photographed with a Polaroid camera and documented using electrophoresis gel documentation system [44].

STATISTICAL ANALYSIS

The data obtained were analysed using SPSS (version 20) statistical software. Standard simple statistical tools such as percentile were used to interpret some of the findings in tabular forms.

RESULTS

The morphological and biochemical characteristic of the isolates revealed that *Staphylococcus aureus* appeared yellow on Mannitol Salt Agar (MSA) with fermentation of sugar indicated by colour variation of the medium and were seen to be Gram-positive cocci during microscopy with biochemical reactions been positive for catalase and coagulase tests. *Escherichia coli* appeared green metallic sheen on Eosin Methylene Blue Agar (EMBA) and were seen to be Gram negative bacilli with biochemical reactions on indole test positive with red surface layer observed while citrate test was negative with no colour variation of the agar slant and Voges-Proskauer negative (Table 1).

Table-1: Cultural, Morphological and Biochemical Characteristics of Isolates from urine samples of patients attending Urology Clinic of DASH Lafia, Nasarawa State, Nigeria

Cultural character	Morphological character		Biochemical character				Inference	
	Gram reaction	Morphology	ind	cit	vp	cat		coag
Pinkish colony on MSA	+	cocci	+	+				<i>S. aureus</i>
Greenish metallic sheen colony on EMBA	-	Rod	+	-	-			<i>E. coli</i>

Key: + = positive, - = negative, ind = Indole test, cit = Citrate test, vp = Voges-Proskauer test, cat =Catalase test, cog = coagulase test

The bacterial count of the isolates in relation to patients analyzed indicated a total number of 202 patients analyzed during the research exercise with high rate of *Staphylococcus aureus* infection (30.69) than *Escherichia coli* infection (20.79). The highest percentage rate recorded was 17 (24.14) in *S. aureus*

and 11 (31.43) in *E. coli*. *Escherichia coli* showed higher resistance value than *S. aureus*. Resistance to the antibiotics analysed was higher in *E. coli* 12 (28.50) than in *S. aureus* (11.29), with a highest resistance rate of 3 (4.69) in *S. aureus* and 4 (9.52) in *E. coli* respectively, as shown on table 2.

Table-2: Bacterial Count of the Isolates in Relation to patients analysed

Patients	Bacterial Isolates (CFU \geq 10 ⁵ /mL)		No. of Resistant isolates (CFU \geq 10 ⁵ /mL)	
	<i>S. aureus</i> (%)	<i>E. coli</i> (%)	<i>S. aureus</i> (%)	<i>E. coli</i> (%)
29	7 (24.14)	7 (24.14)	2 (3.13)	1 (2.38)
40	17 (42.50)	8 (20.00)	3 (4.69)	1 (2.38)
35	12 (34.29)	11 (31.43)	1 (1.56)	4 (9.52)
35	11 (31.43)	8 (22.86)	1 (1.56)	4 (9.52)
25	7 (28.00)	4 (16.00)	-	-
38	8 (21.05)	4 (10.35)	-	-
202	62(30.69)	4 (10.35)	7 (11.29)	7 (11.29)

Key: No. = number, % = percentage, CFU = Colony Forming Unit, mL = millilitres.

Antibiotic Susceptibility among *Staphylococcus aureus* and *Escherichia coli*

The antibiotic susceptibility of *S. aureus* showed that Levofloxacin was most effective against the test organism 46 (74.19%), Norfloxacin been most

intermediate resistance 22(35.48%) and Ampicillin had the most resistance result among the organisms tested 18 (29.03%). Of the total antibiotics analyzed against *E. coli*, Ciprofloxacin had the highest proportion outcome 27 (64.29%) while Amoxycillin/clavulanic acid showed

highest predominance of intermediate result 14 (33.33%). From the analysis, Rifampicin showed the

highest incidence of drug resistance 15 (35.71) against the isolate as shown on table 3.

Table-3: Antibiotic Susceptibility Profile among *Staphylococcus aureus* and *Escherichia coli* isolates

Antibiotics(µg)	Staphylococcus aureus			Escherichia coli		
	Susceptible (%)	Intermediate (%)	Resistant (%)	Susceptible (%)	Intermediate (%)	Resistant (%)
CPX (10)	44(70.96)	6(9.68)	13(20.97)	27(64.29)	1(2.38)	13 (30.96)
LEV (20)	46(74.19)	12(19.35)	5(8.06)	-	-	-
RD (20)	38(61.29)	20(32.26)	4(6.45)	-	-	-
S (30)	39(62.90)	13(20.97)	10(16.13)	26(61.90)	7(16.67)	6(14.29)
NB (10)	27(43.55)	22(35.48)	14(22.58)	-	-	-
E (30)	34(54.84)	21(33.87)	8(12.90)	-	-	-
CN (10)	35(56.45)	19(30.65)	6(9.68)	23(54.76)	13(30.95)	6(14.29)
CH (30)	33(53.23)	17(27.42)	13(20.97)	-	-	-
APX (20)	26(41.94)	19(30.65)	18 (29.03)	-	-	-
AMX (30)	34(54.84)	13(20.97)	16(25.80)	-	-	-
OFX (10)				22(52.38)	8(19.05)	12(28.57)
PEF (10)				15(35.71)	12(28.57)	15(35.71)
NA (30)				19(45.24)	11(26.19)	12(28.57)
AU (30)				22(52.38)	14(33.33)	6(14.29)
SXT (30)				25(59.52)	8(19.05)	10(23.81)
CEP (10)				22(52.38)	9(21.43)	11(26.19)
PN (30)				17(40.48)	11(26.19)	13(30.95)

Key: CPX= ciprofloxacin, LEV = levofloxacin, RD = rifampicin, S = streptomycin, NB = norfloxacin, E= erythromycin, CN = gentamycin, CH = chloramphenicol, APX = ampicillin, AMX = amoxil. CPX = ciprofloxacin, SXT = septrin, PN = ampicillin, OFX = triviid, CEP = ceporex, NA = nalidixic acid, S = streptomycin, PEF = peflacin, CN = gentamycin, AU = Amoxicillin/clavulanic acid, - = not applicable.

Plasmid Variation among Multi-drug Resistant *Staphylococcus aureus* isolates

The plasmid disparity of resistant *S. aureus* isolate showed various plasmids with varying sizes

(bands) ranging from 2500bp to 6000bp. All the isolates presented only one plasmid except for isolate S₁₃⁴ which presented 3 bands with different sizes ranging from 3000bp, 3600bp and 6000bp (Plate 1).



Plate-1: Agarose Gel Electrophoresis of Plasmid DNA from the MDR *Staphylococcus aureus* Isolates

Lane L: Supercoil DNA ladder composed of DNA fragments (in base pairs).

Plasmid Variation among Multi-drug Resistant *Escherichia coli* Isolates

The plasmid disparity of multidrug resistant *E. coli* from this study revealed only one band with

plasmid size of 11, 849bp. The occurrence of Plasmids Resistant among *Escherichia coli* Isolates is shown in Table 7 and agarose gel electrophoresis of plasmid DNA from the MDR *Escherichia coli* I isolates (Plate 2).

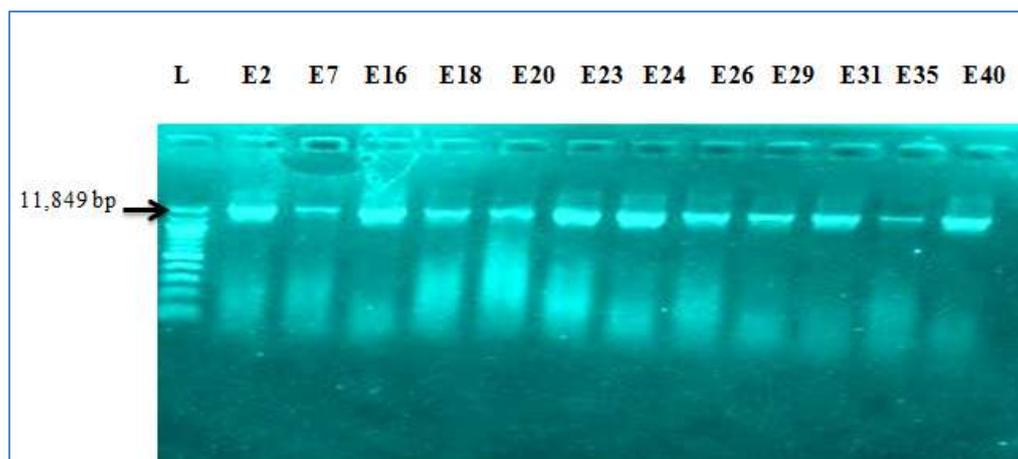


Plate-2: Agarose Gel Electrophoresis of Plasmid DNA from the MDR *Escherichia coli* Isolates

Lane L: Supercoil DNA Ladder Composed of DNA Fragments (In Base Pairs).

DISCUSSION

Urinary tract infection is a major cause of morbidity and mortality. The findings from this work revealed the bacteriuria status of the patients indicated a high rate of *Staphylococcus aureus* infection (30.69%) than *Escherichia coli* infection (20.79%). This could be attributed to contamination of the hand or of the skin surface covering the genitalia. The uropathogens isolated in this research work exhibited high susceptibility rate to Levofloxacin, Ciprofloxacin, and Streptomycin as implicated in *S. aureus* then Ciprofloxacin, Streptomycin, and Septrin implicated in *E. coli*. This result could be as a result of not abusing drugs by the patients or the organisms had not started developing resistance to those antibiotics within the study population. A total number of 19 (18.27%) isolates were recorded resistance for more than two antibiotics (MDR). This is in agreement with the research done by Haruna *et al.*, [45] and Alabi *et al.*, [46]. The high percentage resistance of the isolates in this study is not shocking as it possibly indicates that some of the resistance was as a result of antibiotic misuse or abuse or possibly that the isolates are carrying plasmids for antibiotics resistance as it was also reported by Ngwai *et al.*, [47]. This is also similar to work carried out by Eugene *et al.*, [48] who also documented that resistance to antibiotics by the bacteria might be due to various reasons such as gross misuse of these drugs, over production of target sites, inactivation of the antibiotic, alteration of membrane permeability and spontaneous mutation through DNA transfer amongst others in Uyo city in Akwa Ibom State, Nigeria.

In this research work, the plasmid analysis revealed that multiple drug resistant *S. aureus* isolates harbor one or more plasmids with different molecular weights ranging from 2500 base pair (bp) to 6000bp while multiple resistant *E. coli*. Isolates harbor one single plasmid size type of 11,849bp which denoted that

their resistance is not chromosomal borne. This is in agreement with work carried out by Van Hal *et al.*, [49] in which they documented that MDR plasmids might be acquired by susceptible bacteria through treatment with antibiotics that can induced and hand-picked for horizontal gene transfer. It was also reported by other scientists that the possession of plasmids may be through careful pressure from increased use and misuse of antimicrobial agents [50]. However other scientists assumed that the acquisition of resistance may be due to chromosomal mutations or plasmids that are capable of transferring from one strain of organism to another even across the species in addition to environmental influence. Therefore, the gene coding for antibiotics resistance may perhaps either coded on the plasmid and chromosomal DNA [51].

In this research work, multiple antibiotics resistant *S. aureus* that were resistant before plasmid curing developed became sensitive to the prescribed antibiotics after curing with ethidium bromide. Multiple antibiotics resistant *Escherichia coli* were resistant prior to plasmid curing still retain their resistant to prescribed antibiotics after curing with ethidium bromide. This finding is comparable to the other work carried out by other researchers who reported that plasmid have encoded gene that offers resistance to naturally occurring antibiotics in modest environmental niche [52,53]. Similarly, the findings from this study is also in agreement with the work performed by Ojo *et al.*, [54] which documented that resistance exposed by some bacterial isolates such as *S. aureus*, *Pseudomonas aeruginosa* and *Proteus mirabilis* are mainly plasmid mediated.

Nevertheless, resistances revealed by some of the bacterial isolates such as *Escherichia coli*, and *S. pyogenes* are chromosomal mediated. Several other resistances to bacterial organisms that are not owed to plasmid or chromosome might be owed to efflux pump mechanism [55] or other factors such as mutation of gene encoding ribosomal protein which decrease penetrability of the cell envelope in enteric bacteria

[56]. The bacterial isolates screening with ethidium bromide resultantly propose that the resistance markers were stably lost which is in line with aforementioned studies done by other researchers [57, 58].

CONCLUSION

Staphylococcus aureus and *Escherichia coli* are among the bacteria linked to Urinary Tract Infection within Urology clinic patients of DASH, Lafia, Nasarawa State, Nigeria. There is a fearsome upsurge of infections caused by antibiotic-resistant bacteria. The result from this study showed high susceptibility rate of Levofloxacin against *S. aureus* and Ciprofloxacin against *E. coli* while high resistance rate was recorded in Ampicillin against *S. aureus* and Reflacin against *E. coli* respectively. Appropriate public health policy on prescribing and use of antibiotics must be established and implemented based on current antibiogram results. This Study also disclosed that the genetic basis for antibiotic resistance in the study location is not entirely plasmid mediated but it could either be chromosomal mediated or plasmid mediated.

RECOMMENDATIONS

1. There is the need for proper observation of personal hygiene by individuals such as the use of clean underwear, proper and adequate cleaning after urinating or defecating to help prevent urinary tract infection. There is also the need to wipe front to back and not back to front during cleaning the genital region and the anal openings. This is especially applicable to females.
2. Routine surveillance and monitoring studies should be conducted by government agencies concern as this will help to provide physicians with knowledge about the most effective empirical treatment of UTIs, since drug resistance among pathogens is in circulation in the community and hospital settings.
3. There is the need for collaborative effort between Government agents and private organizations to continuously give public awareness and education on the rationale behind antibiotic use as well as adopting strict national antibiotic policy to regulate the prescription, sale and use of antibiotics in the pharmacies. Indiscriminate prescription and use of antibiotics must be discouraged in both community and hospital settings by Government agencies too.

COMPETING INTERESTS

Authors declared no competing interests.

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