

Original Research Article

Antifungal Activity of *Moringa stenopetala* (Baker f.) Cufod Against *Alternaria helianthi*Prashith Kekuda T.R.¹, Noor Nawaz A.S.², Raghavendra H.L.^{3*}¹Department of Microbiology, S.R.N.M.N College of Applied Sciences, N.E.S campus, Balraj Urs Road, Shivamogga-577201, Karnataka, India²Department of Agricultural Microbiology, UAS, Agriculture College, Dharwad-580005, Karnataka, India³College of Medical and Health Sciences, Wollega University, Post Box No: 395, Nekemte, Ethiopia***Corresponding Author:**

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Abstract: Sunflower is an important oil seed crop in India. The crop is susceptible to various diseases among which leaf blight caused by *Alternaria helianthi* is more destructive. The present study aimed at determining antifungal potential of *Moringa stenopetala* (Baker f.) Cufod (Moringaceae) an indigenous plant to Ethiopia against *A. helianthi* isolated from sunflower leaves. Antifungal activity was determined by Poisoned food technique. The extract was effective and a dose dependent inhibition of mycelial growth of fungus was observed. At extract concentration 1.5mg/ml and higher, an inhibition of >70% was observed. The plant appears promising and the presence of bioactive principles in the extract might have accounted for the antifungal activity. The plant can be used in the management of leaf blight of sunflower.

Keywords: Leaf blight, Sunflower, *Alternaria helianthi*, *Moringa stenopetala*, Antifungal, Poisoned food technique.

INTRODUCTION

Sunflower (*Helianthus annuus* L.; Family Asteraceae) is one among the important oil seed crops in India and other countries. The crop suffers from various diseases such as leaf blight (*Alternaria helianthi*), rust (*Puccinia helianthi*), Powdery mildew (*Erysiphe cichoracearum*), Downy mildew (*Plasmopara halstedii*), Root rot (*Macrophomina phaseoli*), Collar rot (*Sclerotium rolfsii*) and Verticillium wilt (*Verticillium dahliae*). Among these, leaf blight caused by *Alternaria helianthi* (Hansf.) Tubaki and Nishihara is important one. The disease is recognized as potentially destructive in several countries such as India, Yugoslavia, Australia, Uganda and South Africa. In severe cases, the disease results in 70-80% yield loss [1-6]. The disease is destructive and causes reduction in flower size, number of seeds per head, seed weight and oil content. It can often lead to premature defoliation and stem lodge. The disease is characterized by typical dark brown to black, circular to irregular spots on leaves. On stem, petioles, sepals and petals, linear necrotic lesions are observed at different stages of growth. The disease affects the quality of seeds by affecting germination and initial vigour of the seedlings. The disease severity is known to increase or decrease depending on the environmental conditions during crop growth. It is more dangerous in tropical and subtropical regions due to favorable temperature and humidity [1,7,8,9].

Diseases caused by fungi represent the dominant group of plant diseases. These diseases are controlled by the use of synthetic fungicides. Interest in botanicals as antifungal agents is triggered due to drawbacks such as environmental pollution, high cost, development of resistance in fungi and toxicity to non-target organisms associated with inappropriate use of synthetic chemicals [4,10-13]. *Moringa stenopetala* (Baker f.) Cufod (Moringaceae) is an underutilized, fast growing, domesticated *Moringa* species indigenous to southern Ethiopia. It is an important indigenous vegetable food crop and contains high protein content. The seeds are promising as they possess flocculating property which can be exploited in purifying turbid water [14-17]. The plant is used traditionally as food and to treat malaria, hypertension, asthma, diabetes, common cold, wounds, retained placenta and stomach problem [18]. The plant possesses antileishmanial activity [18] and antitrypanosomal activity [19]. The plant is reported to exhibit antimicrobial [20-22], blood pressure lowering [23], anticoccidial [24], antihyperglycemic [25,26] and antihyperlipidemic activity [26]. The objective of the present study was to evaluate antifungal potential of leaf extract of *M. stenopetala* against *A. helianthi* isolated from leaf blight of sunflower.

MATERIALS AND METHODS

Collection and extraction of plant

The plant was collected from Ambo city located 112 km south of Addis Ababa, the capital city of Ethiopia. The plant was authenticated by referring regional flora [27]. The extraction was carried out by maceration process. The leaves were separated, washed well, dried under shade and powdered. 10g of leaf powder was transferred into a flask containing 100ml of methanol. The flask was sealed and left for 48 hours with occasional stirrings. After filtration through Whatman No. 1 filter paper, the filtrate was concentrated at 50°C in an oven. The condensed extract was used for antifungal assay [22].

Isolation of fungus

The fungus *A. helianthi* was isolated from sunflower leaves showing typical symptoms of leaf blight. The surface sterilized leaves were cut into small pieces and placed on sterile Potato dextrose agar (PDA). The plates were incubated at 27±1°C until profuse growth of the fungus was observed. The fungus was then transferred into sterile PDA slant and purified through repeated subculturing. The identification of the isolate was made on the basis of cultural and microscopic characteristics. The pathogenicity test for the isolate was carried out by applying Koch's postulates. The seedlings of susceptible variety (KBSH-44) was inoculated with spore suspension of the isolate and the seedlings were incubated at ambient conditions namely temperature of 27±1°C and relative humidity of 80% for the development of blight symptoms. Reisolation was made from the inoculated seedlings and the isolate completely resembled with the original culture in all respects [5].

Antifungal activity of leaf extract

Poisoned food technique was employed to screen antifungal activity of leaf extract of *M. stenopetala*. Different concentrations of leaf extract (0.1, 1.0, 1.5 and 2.0mg/ml of medium) were used to poison the sterilized and cooled (45°C) PDA medium. The poisoned PDA media were poured into sterile petri dishes and allowed to solidify. 5mm mycelial discs were aseptically cut from 7 days old colony of *A. helianthi* on PDA plates and placed at the centre of control (without extract) and poisoned PDA plates. The plates were incubated at 27±1°C for 5 days in upright position. The diameter of colonies developed on plates was measured in mutual perpendicular directions after incubation.

Antifungal activity of leaf extract (in terms of inhibition of mycelial growth of fungus) was determined using the formula:

$$\text{Inhibition of mycelial growth (\%)} = (A - B / A) \times 100,$$

where 'A' denotes diameter of colony on control plate and 'B' denotes diameter of colony on poisoned plate [22]. The experiment was conducted in triplicates. The results are presented as Mean±Standard Deviation (n=3).

RESULTS AND DISCUSSION

Members belonging to the genus *Alternaria* are cosmopolitan in nature. Species such as *A. alternata*, *A. solani*, *A. macrospora*, *A. porri*, *A. carthami* and *A. helianthi* are known to cause various diseases in their respective hosts [28-31]. *A. helianthi* is recognized as a destructive pathogen of sunflower. It has been shown that the use of chemicals, biocontrol microorganisms and plant extracts have shown to reduce the incidence leaf blight of sunflower and mycelial growth of *A. helianthi* [4,5,32,33]. In the present study, we determined the antifungal effect of leaf extract of *M. stenopetala* against *A. helianthi* by Poisoned food technique. This method is widely used to evaluate antifungal activity of various types of substances including plant extracts. If the sample has antifungal effect then reduction in the mycelial growth occurs in poisoned plate [5,31,34]. Table 1 and Figure 1 show the result of antifungal potential of *M. stenopetala* against mycelial growth of *A. helianthi*. Poisoning of the medium with extract resulted in reduction in mycelial growth of fungus. The inhibitory activity was concentration dependent. More than 70% inhibition of the fungus was observed at concentration 1.5mg/ml and higher. In a similar study, Waghe *et al* [5] showed dose dependent suppression of mycelial growth of *A. helianthi* by certain plant extracts among which marked activity was displayed by neem. In another study, Devi *et al.*[3] screened inhibitory effect of 20 plants against mycelial growth and spore germination of *A. helianthii*. Leaf extract of *Acalypha indica* displayed marked antifungal potential. More recently, we showed concentration dependent antifungal effect of methanol extract of leaf of *M. stenopetala* against three phytopathogenic fungi namely *Colletotrichum capsici*, *Fusarium oxysporum* and *Bipolaris sorokiniana* [22] and seed borne fungi [35].

Table 1: Colony diameter of *A. helianthi* in control and poisoned plates

Concentration	Colony diameter (cm)
0.0mg/ml	5.866±0.057
0.1mg/ml	5.333±0.057
1.0mg/ml	3.966±0.152
1.5mg/ml	1.733±0.115
2.0mg/ml	0.966±0.152

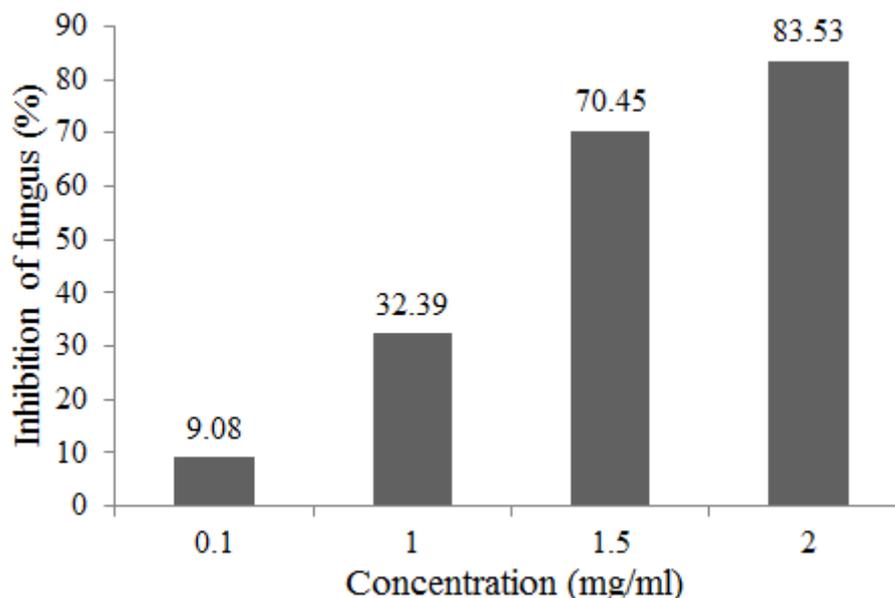


Fig-1: Extent of inhibition of *A. helianthi* by extract

CONCLUSION

Screening plants for antifungal activity against several phytopathogenic fungi as alternate strategy for plant disease control has been intensified nowadays due to problems associated with the use of synthetic fungicides. Botanicals appear to be cheaper and safer alternates for chemical agents as studies have shown the potential of botanicals to inhibit various phytopathogenic fungi. The result of the present study is promising as the leaf extract exhibited marked antifungal potential which may be attributed to the presence of bioactive principles in extract. Further studies are to be conducted to evaluate management of leaf blight in field conditions using *M. stenopetala* leaf.

REFERENCES

- Carson, M. L. (1985). Epidemiology and yield losses associated with *Alternaria* blight of sunflower. *Phytopathology*, 75(10), 1151-1156.
- Amaresh, Y. S., Naragund, V. B., Anahosur, K. H., Kulkarni, S., & Patil, B. V. (2003). Influence of sowing dates and weather factors on development of *Alternaria* leaf blight of sunflower. *Karnataka Journal of Agriculture Sciences*, 16(1), 68-71.
- Devi, A. P., Mohan, S., & Thiribhuvanamala, G. (2013). Antifungal activity of plant extracts against by *Alternaria helianthi*. *Journal of Biopesticides*, 6(2), 231-236.
- Shilpa, M. E., Vikas, H. M., Sowmya, P., Srikantaiah, M., & Dorajeero, A. V. D. (2015). In vitro and in vivo evaluation of antagonistic microorganisms on the incidence of leaf blight of sunflower caused by *Alternaria helianthi*. *Plant Archives*, 15(1), 411-416.
- Waghe, K. P., Wagh, S. S., Kuldhar, D. P., & Pawar, D. V. (2015). Evaluation of different fungicides, bioagents and botanicals against *Alternaria* blight caused by *Alternaria helianthi* (Hansf) of sunflower. *African Journal of Agricultural Research*, 10(5), 351-358.
- Devi, A. P., Mohan, S., Murugapriya, M., Kalieswari, M., & Maharaja, N. (2016). Morphological and cultural characters in determination of virulence of *Alternaria helianthi* on sunflower. *World Journal of Agricultural Sciences*, 12(2), 91-96.
- Calvet, N. P., Ungaro, M. R. G., & Oliveira, R. F. (2005). Virtual lesion of *Alternaria* blight on sunflower. *Helia*, 28, 89-100.
- Basarigid, K., Karuna, K., Shadakshari, Y. G., & Athoni, B. K. (2012). Isolation of toxin produced by *Alternaria helianthi* causing blight in sunflower. *International Journal of Plant Protection*, 5(2), 278-282.
- Basarigid, K., Karuna, K., Jagadish, K. S., Shadakshari, Y. G., & Geetha, K. N. (2013). Evaluation of different culture media for *Alternaria helianthi* causing blight in sunflower. *International Journal of Plant Protection*, 6(2), 457-461.
- Fawzi, E. M., Khalil, A. A., & Afifi, A. F. (2009). Antifungal effect of some plant extracts on *Alternaria alternata* and *Fusarium oxysporum*. *African Journal of Biotechnology*, 8(11), 2590-2597.
- Dellavalle, P. D., Cabrera, D., Alem, D., Larrañaga, P., Ferreira, F., & Rizza, M. D. (2011). Antifungal activity of medicinal plant extracts against phytopathogenic fungus *Alternaria* spp. *Chilean Journal of Agricultural Research*, 71(2), 231-239.
- Nashwa, S. M. A., & Abo-Elyousr, K. A. M. (2012). Evaluation of various plant extracts against the early blight disease of tomato plants under

- greenhouse and field conditions. *Plant Protection Science*, 48, 74–79.
13. Rodino, S., Butu, M., Petrache, P., Butu, A., & Cornea, C. P. (2014). Antifungal activity of four plants against *Alternaria alternata*. *Scientific Bulletin. Series F. Biotechnologies*, 18, 60-65.
 14. Abuye, C., Urga, K., Knapp, H., Selmar, D., Omwega, A. M., Imungi, J. K., & Winterhalter, P. (2003). A compositional study of *Moringa stenopetala* leaves. *East African Medical Journal*, 80(5), 247-252.
 15. Melesse, A., Tiruneh, W., & Negesse, T. (2011). Effects of feeding *Moringa stenopetala* leaf meal on nutrient intake and growth performance of Rhode island red chicks under tropical climate. *Tropical and Subtropical Agroecosystems*, 14, 485-492.
 16. Seifu, E. (2012). Physicochemical properties of *Moringa stenopetala* (Haleko) seeds. *Journal of Biological Sciences*, 12, 197-201.
 17. Seifu, E. (2014). Actual and potential applications of *Moringa stenopetala*, underutilized indigenous vegetable of Southern Ethiopia: A review. *International Journal of Agricultural and Food Research*, 3(4), 8-19.
 18. Mekonnen, Y., & Gessesse, A. (1998). Documentation on the uses of *Moringa stenopetala* and its possible antileishmanial and antifertility effects. *Ethiopian Journal of Science*, 21(2), 287-295.
 19. Mekonnen, Y., Yardley, V., Rock, P., & Croft, S. (1999). In vitro antitrypanosomal activity of *Moringa stenopetala* leaves and roots. *Phytotherapy Research*, 13(6), 538-539.
 20. Walter, A., Samuel, W., Peter, A., & Joseph, O. (2011). Antibacterial activity of *Moringa oleifera* and *Moringa stenopetala* methanol and n-hexane seed extracts on bacteria implicated in water borne diseases. *African Journal of Microbiological Research*, 5(2), 153-157.
 21. Chekesa, B., & Mekonnen, Y. (2015). Antibacterial activity of *Moringa stenopetala* against some human pathogenic bacterial strains. *Science, Technology and Arts Research Journal*, 4(2), 190-198.
 22. Raghavendra, H. L., Kekuda, P. T. R., Vijayananda, B. N., Duressa, D., & Solomon, T. (2016). Nutritive composition and antimicrobial activity of *Moringa stenopetala* (Baker f.) Cufod. *Journal of Advances in Medical and Pharmaceutical Sciences*, 10(3), 1-9.
 23. Mengistu, M., Abebe, Y., Mekonnen, Y., & Tolessa, T. (2012). In vivo and in vitro hypotensive effect of aqueous extract of *Moringa stenopetala*. *African Health Sciences*, 12(4), 545-551.
 24. Meskerem, A., & Boonkaewwan, C. (2013). Protective Effects of *Moringa stenopetala* leaf supplemented Diets on *Eimeria tenella* Infected Broiler Chickens in Debre Zeit, Central, Ethiopia. *Kasetsart Journal (Natural Science)*, 47, 398-406.
 25. Sileshi, T., Makonnen, E., Debella, A., & Tesfaye, B. (2014). Antihyperglycemic and subchronic toxicity study of *Moringa stenopetala* leaves in mice. *Journal of Coastal Life Medicine*, 2(3), 214-221.
 26. Toma, A., Makonnen, E., Mekonnen, Y., Debella, A., & Addisakwattana, S. (2014). Intestinal α -glucosidase and some pancreatic enzymes inhibitory effect of hydroalcoholic extract of *Moringa stenopetala* leaves. *BMC Complementary and Alternative Medicine*, 14, 180.
 27. Verdcourt, B. (1986). Moringaceae. In: Flora of Tropical East Africa. Polhill R.M (Editor). *East African Natural History Society, Kenya*. p 7.
 28. Bashan, Y., Levanony, H., & Or, R. (1991). Association between *Alternaria macrospora* and *Alternaria alternata*, causal agents of cotton leaf blight. *Canadian Journal of Botany*, 69(12), 2603-2607.
 29. Aslam, A., Naz, F., Arshad, M., Qureshi, R., & Rauf, C. A. (2010). In vitro antifungal activity of selected medicinal plant diffusates against *Alternaria solani*, *Rhizoctonia solani* and *Macrophomina phaseolina*. *Pakistan Journal of Botany*, 42(4), 2911-2919.
 30. Singh, G., Gupta, S., & Sharma, N. (2014). In vitro screening of selected plant extracts against *Alternaria alternata*. *Journal of Experimental Biology and Agricultural Sciences*, 2(3), 344-351.
 31. Taware, M. R., Gholve, V. M., & Dey, U. (2014). Bio-efficacy of fungicides, bioagents and plant extracts/botanicals against *Alternaria carthami*, the causal agent of *Alternaria* blight of Safflower (*Carthamus tinctorius* L.). *African Journal of Microbiology Research*, 8(13), 1400-1412.
 32. Mesta, R. K., Benagi, V. I., Kulkarni, S., & Basavarajappa, M. P. (2011). Management of *Alternaria* blight of sunflower through fungicides. *Karnataka Journal of Agricultural Sciences*, 24(2), 149-152.
 33. Reddy, A. B., Sagar, S. D., Mesta, R. K., & Goud, S. I. (2016). Chemical management of *Alternaria* leaf blight of sunflower. *Journal of Farm Sciences*, 29(1), 119-120.
 34. Toubia, E. P., Zakaria, M., & Tahereh, E. (2012). Anti-fungal activity of cold and hot water extracts of spices against fungal pathogens of Roselle (*Hibiscus sabdariffa*) in vitro. *Microbial Pathogenesis*, 52(2), 125-129.
 35. Kekuda, P. T. R., Raghavendra, H. L., Solomon, T., & Duressa, D. (2016). Antifungal and antiradical potential of *Moringa stenopetala* (Baker f.) Cufod (Moringaceae). *Journal of Bioscience and Agriculture Research*, 11(1), 923-929.