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Original Research Article

Variability of Polyphenols, Antioxidant Activity and UFLC Phenolic Acid Profiles of Different Sorghum Genotypes

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Abstract

Sorghum stands as the fifth most valuable global cereal crop, widely grown in semi-arid and arid regions of the world. This present investigation details the variability of polyphenols, scavenging activity in terms of ascorbic acid equivalents of 60 grain sorghum genotypes collected from various countries and were grown at Indian Institute of Millet Research fields which include 38 white, 15 red, and 7 brown pericarp grain sorghum genotypes. Polyphenols range from 575.05 to 3161.87 mg GAE. kg⁻¹, 888.33 to 4230.14 mg GAE.kg⁻¹, 1274.91 to 2885.72 mg GAE. kg⁻¹ in white, red and brown sorghum genotypes respectively. The DPPH radical scavenging activity ranges from 555.74 to 6058.80 mg AAEQ. kg⁻¹, 1190.19 to 6549.59 mg AAEQ.kg⁻¹ and 2174.43 to 5494.40 mg AAEQ. kg⁻¹, in white, red and brown sorghum grain genotypes respectively. Ultra fast liquid chromatography (UFLC) profiling of phenolic acids done in phenolic extracts of red, white and brown sorghum genotypes showed that analytes were derivatives of benzoic acid and cinnamic acid. Total of eleven different phenolic acids were identified, of which ferulic acid is highly expressed in white and red sorghum genotypes only.

Keywords: Totalphenols, Scavenging activity, Polyphenols, Sorghum, Antioxidants, phenolic acid profiling.

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INTRODUCTION

Free radicals are derived form of oxygen and nitrogen, they are effective agents of oxidative stress related to pathogenesis of various degenerative diseases. Oxidative stress reflects an imbalance in favor of free radicals as a result of increase in production or depletion of antioxidant moieties. The Oxidative stress is considered to be the contributory factor in neurodegenerative disorders such as Alzheimer's and Parkinson's diseases and also initiates and promotes the progression of a number of chronic diseases such as diabetes, cancer, cardiovascular diseases, cataract and inflammation. Therefore the intake of phenolic compounds is thought to have more health benefits such as reducing oxidative stress and providing anti-inflammatory, anti-carcinogenic properties [1].

Awareness on natural antioxidants and usage in food products have increased recently because of uncertainty about long term usage of artificial antioxidants such as Butylated hydroxytoluene (BHT) and Butylated hydroxyanisole (BHA). In addition to their long term protection and ability to enhance food quality, these natural antioxidants can also act as good scavengers of free radicals that are produced in

biological systems and provide additional health benefits to consumers. Experiments have suggested that these compounds showed anti radical property thereby involved in various biological functions [2]. Recent studies have suggested the role of plant phenolic compounds as natural antioxidants in foods. Several experimental works have drawn co-relationship between phenolic content and antioxidant property in millet extracts and in processed foods [3]. Some authors found a co-relationship between polyphenols and scavenging activity in plant extracts, while some could not establish any relationship. Cereals contain wide variety of phytochemical agents such as polyphenols, tannins and flavonoids. Among all cereals, maize and red sorghum were found to have high polyphenolic content [4]. Fourteen commonly consumed fresh fruits and ten dry fruits were studied [5] indicated that antioxidant activity and TPC contents of both fresh and dry fruits showed marked variation.

Sorghum bicolor L. Moench is the fifth most important cereal crop in the world after wheat, rice, maize, and barley. Sorghum is grown in semi-arid regions and ranks third in terms of production value, after maize and rice, with a share of 12 % on total

cereal production value (Food and Agriculture Organization of the United Nations Economic and Social Department [6]. In India, the type of sorghum grown is mostly white coloured grain and the white variety of sorghum is widely consumed as staple food and fodder whereas in other countries such as Japan and USA, white sorghum is used only to a small extent and coloured sorghum is grown and used for human and animal consumption.

The phenolic compounds which had the highest antioxidant activity in Shawaya and IS11316 were reported as catechin, 1-O-cafferolglycerol-O-glucoside, taxifolin and pentahydroxyflavanone-(3->4)-catechin-7-O-glucoside [7]. Further the sorghum grain has reported to contain flavonoids, anthocyanins, and tannins. sorghum grain phytochemicals gained lot of interest due to their scavenging activity, nutraceutical property, anti-obese property, preservative property and other health benefits. Hence, sorghum polyphenols could serves as good therapeutic agents [8].

Earlier [9] have compared the scavenging activities of sorghum and sorghum products; they had studied the phenols and scavenging activities of white sorghum in different sub-solvent fractions. In the present study, we report the polyphenols and *in-vitro* scavenging activity of different coloured sorghum flour. The objectives undertaken in the present study were characterization and identification of sorghum genotypes with high scavenging activity, type of phenolic acids and phenolic content and their association in considering it as a functional food.

MATERIALS AND METHODS

Chemicals

UFLC, AR grade solvents and Millipore distilled water (Merck-Millipore, Synergy, UV plus) were used in the extraction and analysis. The chemicals 1,1-Diphenyl-2-picrylhydrazine (DPPH), Ascorbic acid, Phenolic acid standards were procured from Sigma-Aldrich, U.S.A. Folin-Ciocalteau's reagent, Sodium Carbonate, Hydrochloric acid, Gallic acid (3, 4, 5-Trihydroxy benzoic acid) were purchased from Sisco Research laboratories, Mumbai, India. HPLC grade Methanol, Phosphoric acid, and acetonitrile were obtained from Merck India Pvt ltd, Mumbai. Phenolic acid stand UFLC standard compounds were purchased from sigma-Aldrich. All the chemicals were of analytical grade.

Plant Material

Sorghum genotypes (60) were evaluated in augmented design during rabi which include 38 genotypes with white colour pericarp, 15 with red colour pericarp and 7 with brown colour pericarp. These were collected from germplasm pool grown in medium to light soils in the experimental farms of Indian Institute of Millet Research (formerly Directorate of sorghum research), located at

Rajendranagar, Hyderabad, India. The material was sown in three blocks, each with 20 entries. Two elite genotypes, 296 B and C 43 were used as checks in each block. Each genotype was grown in a single line of 5 m length. The material includes germplasm lines belonging to different races from different countries.

Based on the grain pericarp colour genotypes were categorized into three groups, white, red and brown colour grain sorghum genotypes. White color grain sorghum inculudes 27B, 296B, 463B, 7B, AKMS14B, AKR150, AKR354, C43, CSV13, CSV15, IC345194, IC305903, IC305923, I12, IMS9B, IS33648, IS40751, IS4372, RS627, RS673, SPV1258, SPV1293, SPV1471, SPV1606, SPV1616, SPV1731, SPV1732, SPV1733, SPV1734, SPV1750, SPV1760, SPV1775, SPV462. SPV459, SPV711, SPV933. IS18035(originated in India), IS31681(Algeria). Red color sorghum grain and its origin includes IS1212 (China), IS12706, IS20743 (USA), IS16151 (Cameroon), IS20298 (Nigeria), IS23514 (Ethiopia), IS2389, IS3158 (South Africa), IS28141, IS28313, IS12735 (Republic of Yemen), IS29950 (Zimbabwe), IS30538 (Republic of korea), IS4060 (India), IS29241 (Swaziland). Brown color sorghum grain and its origin includes IS12697 (Australia), IS20697, IS715 (USA), IS23992 (Republic of Yemen), IS24462 (South Africa), IS30466 (China), IS30508 (Republic of Korea). The crop was harvested at the panicle (ear head) maturity stage. Panicles were threshed for grain and 250g grain was used for experimental purpose. Whole grain was stored at 4°C for 30 days before extraction.

Extraction of Total Phenols

5g grain samples were ground to fine flour in UDY cyclone sample mill (UDY corporation, USA) to a particle size of 40 μ m separately. One gram of grain flour was mixed with 2.5 ml of methanol with concentrated hydrochloric acid (v/v 99:1) and mixed well in incubator shaker for 1hour at room temperature. The solution was centrifuged at 5000 g for 10 min. The extraction process was repeated twice and the pooled extract was kept at 4° C.

Estimation of Total phenols

Total phenolic content was determined by Folin-Ciocalteu (FC) method which was adapted from [10] with some modifications. The 125 μ L of extract, $400 \mu L$ of methanol, 75 μL of 2 N FCP reagent and 400 μL of 20% Na₂CO₃ were combined in a plastic vial and then mixed well using a vortex. The mixture was allowed to react for 15 min then 1250 μ L of Millipore water was added and mixed well. The solution was centrifuged at 5000 rpm for 15 min in refrigerated centrifuge at $10^0\,\mathrm{C}$ (Thermo Scientific TM Sorvall RC-6 plus, Thermo Scientific corporation, USA). The absorbance was measured at 725 nm using a UV visible spectrophotometer (Model 1601, Corporation, Japan) and the results were expressed in gallic acid equivalents (GAE; mg/kg flour) using a gallic acid (1.25–12.5 µg/mL) standard curve.

Determination of Antioxidant Activity

The antioxidant activity was determined using DPPH invitro assay method [11], 1mL of 100 fold diluted sample extract and 1.5 mL of 0.25 mM DPPH was mixed well. The reaction mixture was then allowed to react for 30 min. The absorbance was measured at 517 nm using a UV visible spectrophotometer (Model 1601,Shimadzu Corporation, Japan) and the results were expressed in ascorbic acid equivalents (AAEQ units/g flour) using ascorbic acid (10-50 μ g/mL) standard curve.

UFLC profiling of phenolic acids

The concentration of specific phenolic acid analysis was carried out with a UFLC system (LC 20AD, Shimadzu corporation, Japan) equipped with a photo diode array (PDA) detector (SPD, M20A, Schimadzu Corporation, Japan). Polyphenols separation was done using a reversed phase C₁₈ column (4.6 x 250 mm, 100 A, 5µm particle size, Phenomenex, India). A gradient binary mobile phase consisting of 0.1 % phosphoric acid (v/v) as solvent A and acetonitrile in solvent B at a flow rate of 1.2 mL/min was used with initial A: B ratio of 90:10 for 13 min, then a linear gradient increase of B solvent to 40% from 13 to 14 min followed by a 3 min hold time and a linear gradient back to the initial A: B ratio of 90:10 in 1 min. Finally an equilibration step with initial concentration for 5 min for a total run time of 22 minutes detection and tentative identification of specific phenolic acid analysis was carried out using the diode array data between 210 to 700 nm along with their retention times and the concentrations were expressed in terms of mg/gm of flour [12].

STATISTICAL ANALYSIS

All experiments were carried out in triplicates and data were reported as mean \pm standard deviation. Statistical software Indostat, version, 2.0 for ANOVA

and Microsoft Excel, 2010 for correlations were used respectively. The differences of mean values among sorghum samples were determined by one-way analysis of variance (ANOVA) followed by Tukey's honestly significant difference (HSD) multiple rank tests at $p \le 0$.05, significance level.

RESULTS AND DISCUSSION

Variability of total phenolic content (TPC) in white, red and brown colour sorghum genotypes

The total phenolic content of 38 white, 15 red, 7 brown pericarp sorghum genotypes was determined and the results were presented in the Tables 1, 2, 3 respectively. Based on the experimental analysis total phenolic content was in the range of 575.05 to 3161.87 mg GAE /kg. The genotype IS31681 had the highest total phenolic content and the AKR150 had the lowest total phenol content. Based on the statistical analysis of the total phenolic content, the genotype IS31681 is significantly different (at CD 1%) from the rest of the white pericarp grains sorghum genotypes tested (Table-1). The present findings are in accordance with [13] who reported a similar range of variation (1.35- 37.73 mg GAE g⁻¹ dm⁻¹) in 224 sorghum samples. [14] have reported that total phenolic content in whole sorghum flour (8.33 mg.g⁻¹), decorticated sorghum flour (1.44 mg.g⁻¹) and sorghum bran (31.95 mg.g⁻¹) of the genotype SC 21. [15] have reported total phenolic content in 36 light colored sorghum lines ranges between 0.60 to 20.73 g GAE kg⁻¹ dm⁻¹. This represents phenolic compounds are quality grade markers for the preparation of several foods due to their enzyme inhibitory activities [16]. The present findings were in agreement with Glennie who reported that concentrations of total phenols of white sorghum grains ranged from 80 to 100 mg/100 g, by [17] is from 109.21 to 116.70 mg/100g.

Table-1: Total phenolic content and DPPH radical scavenging activity of white pericarp sorghum genotypes

			DPPH RSA	
S.No	Genotype	TPC (mg/kg)	(mg AAEQ /kg)	
1	27B	864.9 ± 1.92	5579.06 ± 39.91	
2	296B	814.9 ± 2.27	4073.85 ± 86.11	
3	463B	722.5 ± 3.11	3820.78 ± 98.41	
4	7B	1020.1 ± 3.17	4980.45 ± 34.92	
5	AKMS14B	749.7 ± 0.00	4799.74 ± 293.97	
6	AKR150	575.0 ± 2.95	4471.58 ± 191.90	
7	AKR354	736.6 ± 3.54	4113.53 ± 182.06	
8	C43	677.4 ± 2.67	4748.45 ± 176.95	
9	CSV13	719.2 ± 3.36	5984.71 ± 291.02	
10	CSV15	755.5 ± 3.63	6058.80 ± 186.24	
11	IC 345194	715.6 ± 4.53	5691.94 ± 0.00	
12	IC 305903	1008.3 ± 2.07	5167.85 ± 0.00	
13	IC 305923	913.0 ± 2.07	4759.43 ± 192.45	
14	I12	844.8 ± 7.00	5224.34 ± 35.92	
15	IMS9B	768.8 ± 1.01	4730.20 ± 248.34	
16	IS31681	3161.8 ± 4.08	555.73 ± 156.62	
17	IS33648	1147.6 ± 1.04	4193.61 ± 91.16	

18	IS40751	821.4 ± 3.63	5587.34 ± 167.90
19	IS4372	1068.8 ± 0.07	3874.40 ± 381.67
20	RS627	872.4 ± 3.54	3657.40 ± 0.00
21	RS673	1159.8 ± 0.98	4784.65 ± 218.87
22	SPV1258	1414.5 ± 2.72	5235.85 ± 96.16
23	SPV1293	972.4 ± 4.55	5167.33 ± 0.00
24	SPV1471	924.8 ± 2.70	5481.45 ± 98.13
25	SPV1606	1043.8 ± 1.52	5976.56 ± 106.99
26	SPV1616	634.4 ± 4.79	5268.64 ± 186.24
27	SPV1731	795.4 ± 4.66	3324.33 ± 127.08
28	SPV1732	933.9 ± 3.73	3583.91 ± 155.31
29	SPV1733	843.0 ± 0.49	4078.12 ± 44.71
30	SPV1734	1001.5 ± 6.88	3557.68 ± 970.63
31	SPV1750	773.3 ± 2.72	3711.02 ± 700.60
32	SPV1760	1023.3 ± 1.81	3590.36 ± 92.44
33	SPV1775	857.0 ± 3.45	6017.38 ± 0.00
34	SPV459	926.6 ± 4.43	4290.07 ± 83.84
35	SPV462	876.4 ± 4.15	5943.52 ± 0.00
36	SPV711	1147.6 ± 3.11	4020.65 ± 437.88
37	SPV933	1118.5 ± 5.60	4468.50 ± 64.64
38	IS18035	1108.3 ± 3.54	5392.38 ± 86.60
	C.D. (5%)	57.10	520.25
	C.D. (1%)	76.52	697.22
	C.V. (%)	2.93	5.55
	F (Probability)	0.00	0.00

Results are expressed as mean \pm SD.

The total phenolic content of the red pericarp sorghum genotypes (Table-2) is in the range of 888.33-4230 mg GAE/kg. The genotype IS16151 had the highest total phenolic content followed by IS1212, IS30538 and IS29950, whereas genotype IS28141 showed lowest polyphenols (Table-2). Based on statistical analysis of the TPC, the first four genotypes such as IS16151, IS1212, IS30538 and IS29950 were significantly (at CD 1%) different from each other as well as with the rest of the red pericarp sorghum genotypes tested. Shuyu S *et al.*, [18] have reported the TPC in different sorghum varieties ranging from 174.40

to 1238.83 mg GAE/100 g grain. The other reports supporting the present study showed sorghum with red pericarp at 3.38 mg/g GAE Khoddami *et al.*, 2015, QL33/QL36 (red) 0.88 TPC mg/g GAE, B923296 (red) 0.66 TPC mg/g GAE [7] Generally the total polyphenol content in sorghum is affected by genetic and environmental factors, such as plant color, thickness of the pericarp, and growth conditions. Sorghum with red/purple color has higher total polyphenol contents than tan sorghum grains. The high polyphenol content in red sorghum contributes to a higher resistance to biotic and abiotic stress [19].

Table-2: Total phenolic content and DPPH radical scavenging activity of red pericarp sorghum genotypes

S.No	Conotyno	TPC	DPPH RSA
3.110	Genotype	(mg GAE/kg)	(mg AAEQ /kg)
1	IS1212	3707.8 ± 5.91	6259.32 ± 70.52
2	IS12706	1824.6 ± 2.46	6549.58 ± 0.00
3	IS12735	2078.8 ± 5.91	3566.85 ± 250.83
4	IS16151	4230.1 ± 2.59	1190.18 ± 0.00
5	IS20298	2078.1 ± 8.85	3908.60 ± 367.84
6	IS20743	1668.4 ± 2.36	2678.58 ± 97.69
7	IS23514	2754.1 ± 1.97	2431.30 ± 350.14
8	IS2389	1739.0 ± 4.84	4489.15 ± 208.67
9	IS28141	888.3 ± 2.95	3460.44 ± 40.38
10	IS28313	1072.5 ± 3.48	3517.55 ± 121.15
11	IS29241	3257.8 ± 0.00	2646.73 ± 83.01
12	IS29950	3342.9 ± 5.33	3304.38 ± 232.11
13	IS30538	3597.8 ± 4.99	2096.26 ± 84.72
14	IS3158	1768.8 ± 6.38	3746.02 ± 210.83
15	IS4060	3163.5 ± 5.66	6204.58 ± 146.37
	C.D. (5%)	103.60	367.98

C.D. (1%)	143.79	510.74
C.V. (%)	1.95	4.59
F (Probability)	0.00	0.00

Results are expressed as mean \pm SD.

Similarly, the total phenolic content of the brown pericarp grain sorghum genotypes estimated was in the range of 1274.91-2885.72 mg GAE/kg (Table-3). The genotype IS12697 showed highest total phenolic content, where as IS24462 showed the lowest. As shown in the Table 3 the genotype IS12697 was significantly different from the remaining brown pericarp grain sorghum genotypes tested (at CD 1%). This result is matching with the earlier reports of [7]

who have reported that total phenolic content of brown pericarp IS13116 as 3.58 mg GAE/g. In this study total phenolic content of sorghum genotypes is in the order of Red > white > brown. In addition, [20] investigated de-coated sorghum bran and found that the brown sorghum pericarp variety had higher TPC compared to the black pericarp, indicted the colour of pericarp may not be an ideal indicator of TPC.

Table-3: Total phenolic content and DPPH radical scavenging activity of brown pericarp sorghum genotypes

		TPC	DPPH RSA
S.NO	Genotype	(mg GAE/kg)	(mg AAEQ /kg)
1	IS12697	2885.7 ± 2.86	20174.42 ± 254.93
2	IS20697	1510.35 ± 3.63	2896.39 ± 210.34
3	IS23992	1890.2 ± 0.93	2538.98 ± 95.05
4	IS24462	1274.9 ± 6.60	2929.62 ± 198.25
5	IS30466	1992.3 ± 6.31	4348.99 ± 166.02
6	IS30508	1626.1 ± 3.63	3635.48 ± 670.70
7	IS715	1907.8 ± 5.47	5494.40 ± 159.64
	C.D. (5%)	108.00	707.39
	C.D. (1%)	163.64	1071.79
	C.V. (%)	2.36	8.43
	F (Probability)	0.00	0.00

Results are expressed as mean \pm SD.

DPPH radical scavenging activity

Antioxidants are widely acknowledged for their action against damage by reactive oxygen species, which would be correlated to health beneficial properties such as anti-microbial, reduced oxidative stress, anti-inflammatory and anti-cancer activity [7]. Several methods have been developed including DPPH and ABTS radical scavenging methods. The DPPH radical, is widely used to evaluate the free radical scavenging activity of hydrogen donating antioxidants in many plant extracts [21]. Therefore, antioxidant capacity of sorghum grain was determined by using DPPH assay.

All values of DPPH radical scavenging activity of total phenolic fractions are expressed as milligrams of Ascorbic acid equivalent per gram of grains. The DPPH free radical scavenging activity of white, red and brown pericarp sorghum genotypes is shown in Tables 1, 2, 3 respectively. The results showed that the antioxidant activity of red coloured sorghum genotypes was significantly higher than that of white and brown pericarp sorghum genotypes. This is similar to the gradation of polyphenols Red > white > Brown. The estimated DPPH radical scavenging activity of white pericarp sorghum genotypes is in the range of 555.74-6058.80 mg AAEQ /kg (Table-4). Similarly [18] have reported that white sorghum

genotype such as Longmi sorghum has the DPPH RSA of 920 mg AAEQ/ kg. Also, it was reported that, white sorghum had the antioxidant activity of 14µmol/g (as ABTS) [22]. In the present study genotypes CSV15 and SPV1775 showed highest DPPH radical scavenging activity and genotype IS31681 had lowest. Based on the statistical analysis CSV15, SPV1775, CSV13, SPV1606, SPV462, IC345194, IS40751, 27 B, SPV1471 and IS18035 were found to be not significantly different from each other. However, CSV15 was significantly different from the remaining genotypes tested (at CD 1%) (Table-1).

Similarly in red sorghum genotypes DPPH radical scavenging activity is shown in Table-2. The DPPH RSA estimated is in the range of 1190.19 -6549.59 mg kg⁻¹ (Table-4). Of all the genotypes tested, high DPPH RSA is observed in IS12706, IS1212 and IS4060 and low is noted in IS16151. Based on the statistical analysis of the red pericarp sorghum genotypes IS12706, IS1212 and IS4060 were not significantly different from each other, however statistically significant difference was found from the rest of the genotypes (at CD 1%) (Table-2). In four brewing sorghum varieties, DPPH RSA was reported includes Hongyingzi (19.05)mg AAEQ/g), Hongzhenzhu (5.31 mg AAEQ/g), Dongbei sorghum

(14.94 mg AAEQ/g) and Jiangsu sorghum (1.61 mg AAEQ/g) [18].

Similarly DPPH radical scavenging activity of brown pericarp grain sorghum genotypes is shown in (Table-3), is in the range of 2174.43- 5494.40 mg AAEQ kg⁻¹ (Table-4). Of all the genotypes tested high DPPH RSA is observed in IS715 followed by IS30466 and low was noted in IS12697. Based on the statistical analysis of the brown pericarp sorghum genotypes IS715 is significantly different from the rest of the

genotypes tested (at CD 1%) (Table-3). In earlier literature it was found that non-pigmented sorghum showed relatively radical scavenging activity ranged from 7% to 67% [23], antioxidant capacity measured by ABTS assay of non-tannin sorghum grains ranged from 9.7–78.9 µmol TE/g sample [24]. To our knowledge, this data will add more information to the existing knowledge on the polyphenols, antioxidant activity, phenolic acid profiles of 60 different pericarp sorghum genotypes across the world, which is important for the selection of sorghums to be used for functional foods.

Table-4: Mean values and range of total phenolic content and scavenging activity of white, red and brown pericarp sorghum genotypes

	8 J F	
	Total phenolic content (mg GAE/kg)	DPPH RSA (mg AAEQ /kg)
White sorghum		
Mean±SD	960.91 ± 405.77	4628.56 ± 1063.27
Range	575.05 - 3161.87	555.74 - 6058.80
SE	65.87	6.16
Red sorghum		
Mean±SD	2478.22 ± 1024.41	3736.62 ± 1569.59
Range	888.33 - 4230.14	1190.19-6549.59
SE	264.7	104.63
Brown sorghum		
Mean±SD	1726.03 ± 262.92	3431.9 ± 1158.59
Range	1274.91 - 1992.39	2174 - 5494.40
SE	99.59	165.51

UFLC Identification and quantification of phenolic acids in white, red and brown pericarp grain sorghum genotypes

Sorghum grain contains phenolic acids, which are located in the pericarp, testa, aleurone layer and endosperm [25]. Sorghum rich in phenols with antioxidant properties is important to identify the individual phenolic compounds, which could allow the sorghum genotype selection and grains production system to provide help in health, food, nutraceutical and pharmaceutical applications [26].

White, red and brown pericarp sorghum genotypes with high phenolic content were selected and analysed for phenolic acids through UFLC. The analysis by UFLC-DAD acidified methanolic extracts revealed the presence of different classes of phenolic acids. The UFLC profile of all these 11 phenolic compounds when taken together showed the same sequence of elution using this method and were confirmed by comparison of retention times, UV spectral data with those of commercial standards (Figure 1a, 1b, 1c). Chromatogram of 11 standard phenolic compounds showing namely gallic acid (5.19 min), salicin (7.8 min), catechin (10.77 min), 4-hydroxy benzoic acid (10.91 min), vanillic acid (13.49 min), chlorogenic acid (12.73), caffeic acid (15.2 min), syringic acid (16 min), ferulic acid (18.68), salicylic

acid (19.57 min) and cinnamic acid (21.5 min) which were eluted at different retention times. Each phenolic acid identified in white, red and brown pericarp grain sorghum genotypes are quantified and presented in Table 6a, 6b and 6c respectively.

Among the 38 white sorghum genotypes only eleven genotypes IS31681, SPV1258, RS673, SPV711, IS 33648, SPV 1732, IC 305923, IS40751, 296B, IMS 9B, IC 345194 rich in total phenolic content were selected and quantified for individual phenolic acids (table 5a). In white sorghum genotypes gallic acid (0.64-76.6 mg/100g), vanillic acid (0.15-5.5)mg/100g), chlorogenic acid (2.22 to 6.64 mg/ 100g), ferulic acid (1.7 to 7.52 mg/100g), salicylic acid (8.95 to 32.2 mg/100g), catechin (2.55 to 9.2 mg/100g) and caffeic acid (0.56 to 18.44 mg/ 100g) were identified. Higher content of gallic acid was detected only in SPV711 genotype, whereas, vanillic acid was detected in IS33648, chlorogenic acid in SPV1258, Ferulic acid in IC305923, salicylic acid in IS31681, catechin was detected only in SPV1258 and caffeic acid was detected only in SPV1258 (Table-6a). Similarly, White pericarp sorghum variety CS3541 was reported to have the phenolic compounds such as gallic acid, phydroxybenzoic acid, vanillic acid, protocatechuic acid, coumaric acid, caffeic acid, ferulic acid and cinnamic acid [19].

Table-6a: Phenolic acids quantified in methanolic extracts of white pericarp grain sorghum genotypes (by elution order)

				/				
Genotype	Gallic acid	Vanillic acid	Chlorogenic acid	Ferulic acid	Salicylic acid	Catechin	Caffiec acid	Total phenolic
								acids
								(mg/100g)
IS31681	n.d.	n.d.	n.d.	n.d.	32.2 ± 0.14	n.d.	n.d.	32.2

SPV1258	12.3 ± 0.42	n.d.	6.64 ± 0.19	n.d.	n.d.	9.2 ± 0.04	18.44 ± 0.33	46.58
RS673	0.63 ± 0.00	0.15 ± 0.00	n.d.	n.d.	n.d.	2.55 ± 0.01	n.d.	3.33
SPV711	76.6 ± 0.42	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	76.6
IS33648	n.d.	5.5 ± 0.28	n.d.	7.45 ± 0.3	n.d.	n.d.	n.d.	12.95
SPV 1732	n.d.	1.5 ± 0.4	2.85 ± 0.06	3.61 ± 0.4	n.d.	n.d.	n.d.	7.96
IC 305923	0.64 ± 0.04	0.42 ± 0.03	n.d.	7.52 ± 0.1	n.d.	n.d.	0.56 ± 0.02	9.14
IS40751	n.d.	0.63 ± 0.04	2.22 ± 0.16	4.06 ± 0.08	8.95 ± 0.07	n.d.	n.d.	15.86
296B	n.d.	0.74 ± 0.05	n.d.	1.8 ± 0.14	n.d.	n.d.	n.d.	2.54
IMS 9B	n.d.	1.2 ± 0.28	5.3 ± 0.13	1.77 ± 0.17	n.d.	n.d.	n.d.	8.28
IC 345194	n.d.	1.85 ± 0.07	6.26 ± 0.19	3.62 ± 0.38	n.d.	n.d.	n.d.	11.73

Values are calculated from the area obtained for each compound and for each authentic standards. Results are expressed as mean \pm SD, n.d-notdetected.

Among the 15 red pericarp sorghum genotypes only eight genotypes rich in total phenolic content, which includes IS1212, IS16151, IS28313, IS29950, IS29241, IS3158, IS30538 and IS4060, were quantified for phenolic acids through UFLC. In red pericarp sorghum genotypes the phenolic acids detected were gallic acid (7.4 mg/ 100g), salicylic acid (28.67 mg/100g), protocatechuic acid (12.85 mg/100g), salicin (1.55 to 44.55 mg/100g), vanillic acid (0.20 to 1.21 mg/ 100g), chlorogenic acid (0.82 to 4.5 mg/100g), ferulic acid (2.6 to 49.7 mg/ 100g) and catechin (1.05 mg/ 100g) (Table-6b). This holds true with the earlier reports where red sorghum variety SC0630 was reported to have the phenolic compounds such as gallic p-hydroxybenzoic acid, vanillic protocatechuic acid, coumaric acid, caffeic acid, ferulic acid and cinnamic acid [19] Of all the phenolic acids observed, only ferulic acid was identified at higher concentration compared to other phenolic compounds in the red grain sorghum genotypes, of which IS29950 showed high content of t-ferulic acid (49.75 mg/ 100g flour). which is akin to the earlier reports where

phenolic acids in the red sorghum variety PAN 3860 were identified include ferulic acid as the major compound followed by protocatechuic acid, protocatechuic aldehyde, caffeic acid and p-coumaric acid [27].

Similarly high total phenolic content containing brown pericarp grain sorghum genotypes, which includes IS12697, IS30466, IS715, IS23992 and IS30508, were evaluated for phenolic acids. The identified phenolic acids were gallic acid (2.85 to 11.58 mg/ 100g), 4-hydroxy benzoic acid (0.57 mg/ 100g), vanillic acid (0.45 mg/ 100g), salicylic acid (0.94 mg/ 100g) and catechin (25.5 mg/ 100g) (Table -6c). High phenolic acid content was noted in IS30508 (32.03 mg/ 100g) followed by IS30466 and IS715 (11.3 mg/ 100g). Similar conclusions have been made in brown color sorghum variety (SC0719) was reported to have the phenolic compounds such as gallic acid, phydroxybenzoic acid, vanillic acid, protocatechuic acid, coumaric acid, caffeic acid, ferulic acid and cinnamic acid [19].

Table-6b: Phenolic acids quantified in methanolic extracts of red pericarp sorghum genotypes (by elution order)

Genotype	Gallic	Salicylic	Protocatechuic	Salicin	Vanilic	Chlorogenic	Ferulic acid	Catechin	Total
	acid	acid	acid		acid	acid			phenolic acids mg/100g
IS16151	n.d.	28.67 ± 0.43	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	28.67
IS1212	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	1.05 ± 0.09	1.05
IS30533	7.41 ± 0.02	n.d.	12.85 ± 0.07	n.d.	n.d.	n.d.	n.d.	n.d.	20.26
IS29950	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	49.75 ± 0.21	n.d.	49.75
IS29241	n.d.	n.d.	n.d.	1.55 ± 0.35	n.d.	n.d.	2.60 ± 0.28	n.d.	4.15
IS4060	n.d.	n.d.	n.d.	44.55 ± 0.49	n.d.	n.d.	n.d.	n.d.	44.55
IS28313	n.d.	n.d.	n.d.	n.d.	1.21 ± 0.01	4.5 ± 0.28	10.65 ± 0.07	n.d.	16.36
IS3158	n.d.	n.d.	n.d.	n.d.	0.20 ± 0.02	0.82 ± 0.05	2.65 ± 0.21	n.d.	3.67

Values are calculated from the area obtained for each compound and for each authentic standards. Results are expressed as mean $\pm SD$, n.d. -not detected

Table-6c: Phenolic acids quantified in methanolic extracts of brown pericarp grain sorghum genotypes

Genotype	Gallic acid	4-Hydroxy benzoic acid	Salicylic acid	Catechin	Vanillic acid	Total phenolic acids (mg/100g)
IS12697	2.85 ± 0.07	n.d.	n.d.	n.d.	n.d.	2.85
IS30466	11.58 ± 0.40	n.d.	n.d.	n.d.	n.d.	11.58
IS715	6.88 ± 0.14	0.57 ± 0.01	0.94 ± 0.04	n.d.	n.d.	8.39
IS23992	ND	n.d.	n.d.	n.d.	0.45 ± 0.07	0.45
IS30508	6.78 ± 0.26	n.d.	n.d.	25.25 ± 0.77	n.d.	32.03

n.d. – not detected.

Values are calculated from the area obtained for each compound and for each authentic standards. Results are expressed as mean $\pm\,S$

The chromatograms are given for the genotype SPV1258 (white sorghum) rich in caffeic acid (RT 12.9 min), IS29950 (red sorghum) rich in ferulic acid (RT 18.68 min) and IS30466 (brown sorghum) rich in gallic acid (RT 3.5 min) (Figure 1a, 1b, 1c). Ferulic acid has been reported as the most abundant bound phenolic compound in sorghum [25] and other cereals. The

contents of ferulic acid in bound fractions in the investigated sorghum grains also showed the highest level compared to other phenolic acids, accounted for over 60% of the total identified phenolic acids and ranged from 1.55 to 85.98 mg/100 g grain (DW). The concentration of ferulic acid (86.84 mg/ 100g) in black gain sorghum was higher than the reported levels in sorghum grains, oat, rice, wheat and barley (8,17,23). Total concentrate caffeic acid varied from 3.49 to 8.17 mg/100 g grain [18]. Similar levels were reported before in other varieties of sorghums [28].

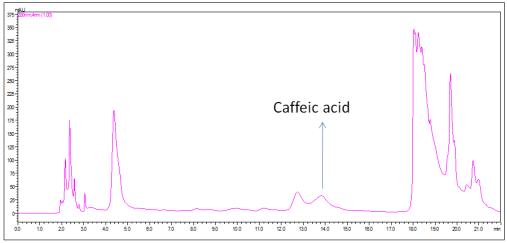


Fig-1a: UFLC Chromatogram of SPV1258 white sorghum genotype rich in caffeic acid eluted at RT 12.9 min

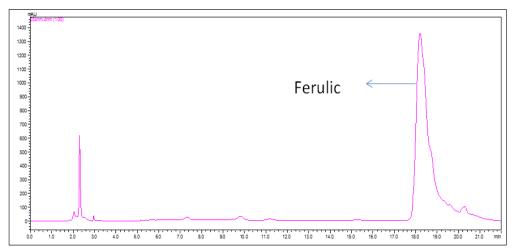


Fig-1b: UFLC Chromatogram of IS29950 red sorghum genotypes ferulic acid eluted at RT 18.68 min

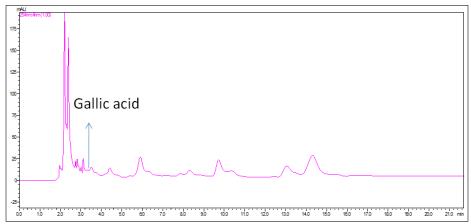


Fig-1c: UFLC Chromatogram of IS30466 brown sorghum genotype rich in gallic acid eluted at RT 3.5 min

Sorghum is a good source of phenolic compounds. Gallic acid was present only in the bound form where as protocatechuic, p-hydroxybenzoic, caffeic, p-coumaric and ferulic acids were found in the free and bound forms. However, cinnamic and vanillic acids were found in the free and/or bound forms only in some sorghum varieties. Two sorghum genotypes namely white (PR6E6) and red (PR6E14) showed four phenolic acids (caffeic, p-coumaric, ferulic and sinapic acids) [29]. In general phenolic acids are bioaccessible. they have bioactivities and serve as potential natural sources of antioxidants in food and biological systems. Earlier literature have suggested that phenolic acids such as ferulic acid, cinnamic acid, p-hydroxybenzoic acid play a major role in inhibiting the growth of several fungal species [30].

Antioxidant capacities and their correlation with phenolic compounds

A negative correlation was found between poly phenolic content and DPPH Radical scavenging activity in white sorghum (r value being -0.589), red sorghum (r value being -0.202) and brown sorghum (r value being -0.111) (Table-5). This result is in accordance with the reports of [31] who did not find a correlation between phenols and antioxidant activity, simply because they used only white sorghums, which are quite low in phenols. However these findings are in contrast to those of the earlier reports where the total phenol content of sorghums is significantly correlated with antioxidant activity [16, 19, 32].

Table-5: Correlation between Total phenolic content and DPPH radical scavenging activity of white, red and brown pericarp sorghum genotypes

i brown pericarp :	sorgnum genoty
	TPC
White sorghum	
Polyphenols	1
DPPH	-0.589924309
Red sorghum	
Polyphenols	1
DPPH	-0.202780306
Brown sorghum	
TPC	1
DPPH	-0.111554331

CONCLUSION

Comparison of the differences in total phenolic content and antioxidant capacities of different varieties of sorghum grains are reported in the present study. Red pericarp sorghums grains showed high phenolic as well as in vitro antioxidant activity. All of the sorghum grains contained high contents of phenolic acids, especially ferulic acid. It is therefore important to consider the genotype while selecting sorghums for human food and animal feed so as to obtain maximum energy and protein availability and they can be used as high antioxidant value-added health foods having nutraceutical or pharmaceutical applications. The results presented in this study will be of significant use to cereal producers and consumers as phenolic acids are known to influence the organoleptic properties and health benefits of whole grains.

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