

Metal Toxicity in Radish Plant with Carotenoids, Chlorophylls Study Under Stress Condition's

Muhammad Ehsan Haider¹, Muhammad Ihsan^{2*}, Muhammad Riaz¹, Muhammad Saleem¹, Gul E Zahra², Lyeba Shoaib², Nafeesa Shehzadi², Asad Shehzaib³

¹Department of Botany, University of Agriculture, Faisalabad, Pakistan

²Institute of Botany, University of Punjab, Lahore Pakistan

³Institute of Horticultural Sciences, UAF, Pakistan

DOI: [10.36348/sijap.2021.v04i7.003](https://doi.org/10.36348/sijap.2021.v04i7.003)

| Received: 07.06.2021 | Accepted: 09.07.2021 | Published: 15.07.2021

*Corresponding author: Muhammad Ihsan

Abstract

Radish is consumed and cultivated globally and is reflected as a part of human-diet, although this root vegetable is not common in many populations. Different varieties of radish have the different soil and climate requirements for better growth and yield but the most beneficial and important agro-technique is nutrition for better performance. For better quality and optimal radish production, fertilization by organic, inorganic and bio-fertilizers are very important constituents. Other pharmaceutical uses of radish include treatment of chronic diarrhea, neuralgic-headache and sleeplessness. Toxicity of Ni in radish plant is confirmed by inhibition of growth, chlorosis, necrosis and wilting. This report based study conducted to investigate the biological, physiological effects of heavy metals on the radish. In this study it was noticed that nickle sulphate act as suppressor on growth of the plant and it also decreased the content of carotenoids and chlorophyll contents such as Chlorophyll-a, b. Statistical analysis was achieved after collection of data by using appropriate computer software such as ANOVA.

Keywords: Carotenoids, chlorophyll a, Chlorophyll b, radish, metals, toxicity, physiological features.

Copyright © 2021 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

INTRODUCTION

Radish is an important and very prevalent root vegetable crop cultivated in the tropical, sub-tropical and in the temperate areas of world. Radish is cultivated both as an annual and the biennial root vegetable crop depends on the purpose of which it is cultivated. It is mainly a cool period crop. However, Asiatic kinds can withstand high temperatures than the varieties of Europe. In mild weather, it can be cultivated almost the whole year excluding few summer months [1, 2].

Eating vegetables aid in inhibiting of many diseases, therefore it is also called as protecting food. Vegetables are the richest, natural and inexpensive source of very protective food that provides fats, minerals, carbohydrates, vitamins and proteins [3]. According to the suggested diet proposal, vegetables feeding should be 300g per day/person. Out of these 300g, 125g the leafy-green-vegetables, 100g deep-rooted-vegetables and the 75g other included. During the metabolism in radish the enzyme myrosinase leads to the formation of isothiocyanates by enzymatic hydrolyses of glucosinolates. The cells of some plants

also pretentious with the cancer due to the abnormal cells proliferation. Bacterial micro-flora, present in the colon of human, also exhibits same enzyme activity [4].

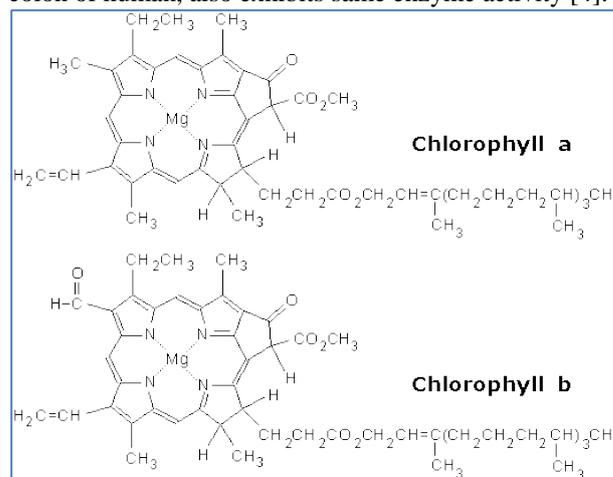


Fig-1: Shows the structures of chlorophyll a and b

Other pharmaceutical uses of radish include treatment of chronic diarrhea, neuralgic-headache and sleeplessness. The whitish color ash is obtained from

the roots processing that act as a therapy for diuresis and constipation [10]. Leaves of Plants contain distinct pores that are called stomata. Endorsement of various heavy-metals by the plants change the mechanisms of growth by triggering secondary responses such as oxidative-damage, may be by accumulation of hydrogen per oxide which occurred by changing balance of ROS (reactive oxygen species) metabolisms. Interaction of metals with the metabolism and growth has communal effects which finally disturbs the growth of plant. Plant built nutraceuticals products are utilized in pharmacological industries. For example, quantity of the ne metal may distress negatively or positively to the other element, either it transform or diminishes the poisonous effects of the heavy-metals [11].

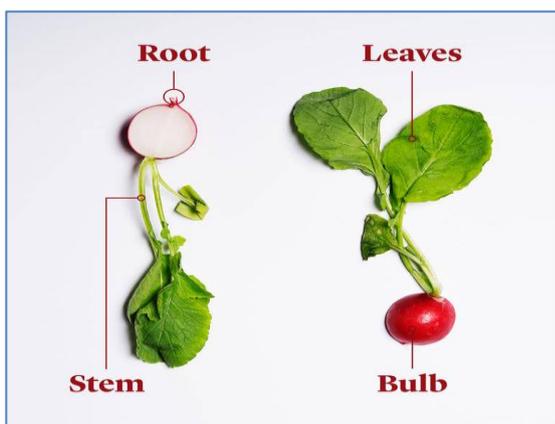


Fig-2: Shows the botanical features of radish plant

This report based study conducted to carry the effects of heavy metals on the radish.

MATERIALS AND METHODS

This report based study conducted to carry the effects of heavy metals on the radish. In this study it was noticed that nickle sulphate act as suppressor on growth of the plant and it also decreased the content of carotenoids and chlorophyll contents (Chlorophyll-a, b). Statistical analysis was achieved after collection of data by using appropriate computer software such as ANOVA [14]. All morphological changes were observed to collect the data for the biochemical analysis. A marked reduction in chlorophyll a and chlorophyll b was noticed with slight elevation of carotenoid.

To evaluate the amount of carotenoids and chlorophyll a and chlorophyll b, methodology. 0.10g of leaf samples were collected through cutting and socked them into a beaker containing 10ml of 80% acetone then these sample were centrifuged after 24 hours at 10,000 rpm for 5min and then spectrophotometer was used to check the supernatant's absorbance 480, 645 and 663 nm[15].

STATISTICAL ANALYSIS

Statistical analysis performed to this data of report and organized in the form of tables.

RESULTS AND DISCUSSIONS

Table-1: Shows the values of ANOVA table of chlorophyll a,b and carotenoids

SOV	DF	Chlorophyll a	Chlorophyll b	Carotenoids
Treatment	3	3.572	4.8299**	9.2***
Error	8	7.568	9.5843	2.2279

The analysis of collected data from radish's chlorophyll a that were grown under controlled conditions is presented in table. Nickle sulphate act as suppressor that bring the reduction in growth as well as chlorophyll a($P \geq 0.001$). Under controlled conditions maximum reduction was observed as compared at 150mM Nickel sulphate that was applied in Foliar medium [16].

The analysis of collected data from radish's chlorophyll b that were grown under controlled conditions is presented in table. Nickle sulphate acts as suppressor that brings the reduction in growth as well as chlorophyll b. Under controlled conditions maximum reduction was observed as compared at 30mM Nickel sulphate that was applied in Foliar medium[17, 18]. Analysis of variance of data for carotenoids of Radish under foliar application of $NiSO_4$ is presented in table observed in carotenoids [19-21].

Toxicity of Ni in radish plant is confirmed by inhibition of growth, chlorosis, necrosis and wilting.

The plants that grow in polluted soil generally have higher heavy metals level depending upon total meditation of soil and the genotype plants. Catalase and heme with increased concentration of Ni may show intrusion in the metabolism of iron in plants. With the aggregate level of nickel the superoxide dismutase and ribonuclease activities and the level proline also increased [12, 13].

The young roots are consumed as raw in salad and prepared as vegetable. It has pungent taste and reflected as an appetizer. The leaves are eaten and cooked as vegetable [5]. The provisions of *Raphanus sativus* are beneficial for gall bladder and the liver troubles. The leaves, flowers, pods and roots play vital role against the urinary complaint, gastrodynia, piles and gram positive bacteria. The salts are taking out from root and are dried then burnt to whitish ash and are utilized to diminish stomach difficulties [6].

Mainly, the growth and yield of vegetable crops depends upon the soil and favorable weather. The

barcodes of plant DNA are utilized for identification of species. The heavy metals prove very toxic for the vegetable crops. They reduce the performance of crops. Many of the plants suffered by the toxicity of Ni [7]. Different varieties of radish have the different soil and climate requirements for better growth and yield but the most beneficial and important agro-technique is nutrition for better performance. The requirements of radish plant fluctuate with soil fertility, agro climatic conditions, soil and humidity [8, 9].

CONCLUSION

Consistent amount of nickel was applied. In this study it was noticed that nickel sulphate act as suppressor on growth of the plant and it also decreased the content of carotenoids and chlorophyll contents and Chlorophyll a, b. Determination of chlorophylls and carotenoids are important to investigate the physiological properties of plants because these are important to help the different features of disease causing microbes and metals.

REFERENCES

1. Agrawal, A. A. (1999). Induced responses to herbivory in wild radish: effects on several herbivores and plant fitness. *Ecology*, 80(5), 1713-1723.
2. Hamza, M. A., & Aylmore, L. A. G. (1992). Soil solute concentration and water uptake by single lupin and radish plant roots. *Plant and soil*, 145(2), 197-205.
3. Shokohifard, G., Sakagami, K., Hamada, R., & Matsumoto, S. (1989). Effect of amending materials on growth of radish plant in salinized soil. *Journal of plant nutrition*, 12(10), 1195-1214.
4. Khan, D. H., & Frankland, B. (1983). Effects of cadmium and lead on radish plants with particular reference to movement of metals through soil profile and plant. *Plant and soil*, 70(3), 335-345.
5. Yildirim, E. R. T. A. N., Turan, M. E. T. I. N., & Donmez, M. F. (2008). Mitigation of salt stress in radish (*Raphanus sativus* L.) by plant growth promoting rhizobacteria. *Roumanian Biotechnol Lett*, 13, 3933-3943.
6. Curtis, I. S. (2003). The noble radish: past, present and future. *Trends in Plant Science*, 8(7), 305-307.
7. El-Beltagi, H. S., Mohamed, A. A., & Rashed, M. M. (2010). Response of antioxidative enzymes to cadmium stress in leaves and roots of radish (*Raphanus sativus* L.). *Notulae Scientia Biologicae*, 2(4), 76-82.
8. Lichtenthaler, H. K., Burkard, G., Kuhn, G., & Prenzel, U. (1981). Light-induced accumulation and stability of chlorophylls and chlorophyll-proteins during chloroplast development in radish seedlings. *Zeitschrift für Naturforschung C*, 36(5-6), 421-430.
9. Lichtenthaler, H. K., Kuhn, G., Prenzel, U., & Meier, D. (1982). Chlorophyll- protein levels and degree of thylakoid stacking in radish chloroplasts from high-light, low-light and bentazon- treated plants. *Physiologia Plantarum*, 56(2), 183-188.
10. Silva, F. B., Costa, A. C., Alves, R. R. P., & Megguer, C. A. (2014). Chlorophyll fluorescence as an indicator of cellular damage by glyphosate herbicide in *Raphanus sativus* L. plants. *American Journal of Plant Sciences*, 2014.
11. Chaparzadeh, N., & Hosseinzad-Behboud, E. (2015). Evidence for enhancement of salinity induced oxidative damages by salicylic acid in radish (*Raphanus sativus* L.). *Journal of Plant Physiology & Breeding*, 5(1), 23-33.
12. Ramakrishna, B., & Rao, S. S. R. (2015). Foliar application of brassinosteroids alleviates adverse effects of zinc toxicity in radish (*Raphanus sativus* L.) plants. *Protoplasma*, 252(2), 665-677.
13. Sun, B. Y., Kan, S. H., Zhang, Y. Z., Deng, S. H., Wu, J., Yuan, H., ... & Li, Y. W. (2010). Certain antioxidant enzymes and lipid peroxidation of radish (*Raphanus sativus* L.) as early warning biomarkers of soil copper exposure. *Journal of Hazardous Materials*, 183(1-3), 833-838.
14. Monterumici, C. M., Rosso, D., Montoneri, E., Ginepro, M., Baglieri, A., Novotny, E. H., ... & Negre, M. (2015). Processed vs. non-processed biowastes for agriculture: effects of post-harvest tomato plants and biochar on radish growth, chlorophyll content and protein production. *International journal of molecular sciences*, 16(4), 8826-8843.
15. Pawłowska, B., Telesinski, A., Płatkowski, M., Streck, M., Snioszek, M., & Biczak, R. (2017). Reaction of spring barley and common radish on the introduction of ionic liquids containing asymmetric cations to the soil. *Journal of agricultural and food chemistry*, 65(23), 4562-4571.
16. Anuradha, S., & Rao, S. S. R. (2009). Effect of 24-epibrassinolide on the photosynthetic activity of radish plants under cadmium stress. *Photosynthetica*, 47(2), 317-320.
17. Lichtenthaler, H. K., Prenzel, U., & Kuhn, G. (1982). Carotenoid composition of chlorophyll-carotenoid-proteins from radish chloroplasts. *Z Naturforsch 37c*, 10-12.
18. Grumbach, K. H., & Britton, G. (1983). Distribution of carotenoids in sub-cellular and sub-plastidic fractions of radish seedlings (*Raphanus sativus*) grown in the presence of bleaching herbicides. *Phytochemistry*, 22(9), 1937-1940.
19. Suzuki, Y., & Shioi, Y. (2004). Changes in chlorophyll and carotenoid contents in radish (*Raphanus sativus*) cotyledons show different time courses during senescence. *Physiologia Plantarum*, 122(2), 291-296.
20. Park, C. H., Park, S. Y., Park, Y. J., Kim, J. K., & Park, S. U. (2020). Metabolite Profiling and Comparative Analysis of Secondary Metabolites in Chinese Cabbage, Radish, and Hybrid *xBrassicoraphanus*. *Journal of Agricultural and Food Chemistry*, 68(47), 13711-13719.
21. Bach, T. J., & Lichtenthaler, H. K. (1983). Inhibition by mevinolin of plant growth, sterol formation and

pigment accumulation. *Physiologia plantarum*, 59(1),

50-60.