

## Application of Oxazole and Oxazolopyrimidine as New Effective Regulators of Oilseed Rape Growth

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**Abstract:** The elaboration of new effective and ecologically friendly regulators improving growth and increasing yield of oilseed rape is an actual problem for modern agriculture. Our work was devoted to screening of new effective plant growth regulators among chemical low molecular weight heterocyclic compounds, derivatives of oxazole and oxazolopyrimidine to improve the germination of seeds and growth of seedlings of oilseed rape (*Brassica napus* L.) of cultivar Kalinivsky. As a result of the conducted experiments, the most effective synthetic compounds that showed a high stimulating effect when used in concentration  $10^{-9}$ M on the growth of 21<sup>st</sup>-day-old oilseed rape seedlings were selected. It was found that biometric indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of derivatives of oxazole and oxazolopyrimidine were increased by an average to 11 - 30 % – by length of shoots, by an average to 8 - 68 % – by total number of roots, and by an average to 5 - 43 % – by total length of roots, as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or on the  $10^{-9}$ M solution of plant hormones auxins IAA (1*H*-Indol-3-ylacetic acid) and NAA (1-Naphthylacetic acid). The content of photosynthetic pigments in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of derivatives of oxazole and oxazolopyrimidine was increased by an average to 14 - 20 % – by content of chlorophyll a, by an average to 15 - 21 % – by content of chlorophyll b, by an average to 16 - 18 % – by content of chlorophyll a+b, as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) and were increased by an average to 14 - 26 % – by content of carotenoids as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or grown on the  $10^{-9}$ M solution of IAA and NAA, respectively. The obtained results confirmed the possibility of using of derivatives of oxazole and oxazolopyrimidine as new effective regulators to improve oilseed rape growth.

**Keywords:** oilseed rape, growth regulators, oxazole and oxazolopyrimidine, auxins IAA and NAA.

### INTRODUCTION

Oilseed rape (*Brassica napus* L.) is an important strategic energy and food crop cultivated in the economically developed countries including the USA, Canada, European Union, China, India, and Australia [1-9]. Oilseed rape is used for the production of edible vegetable oils, animal feed, and biodiesel [2, 10, 11].

Oilseed rape oil is useful for human nutrition for protection of cardiovascular diseases due to lower content of cholesterol and high content of most important cardioprotective substances including unsaturated fatty acids such as linoleic acid (21 %) and alpha-linolenic acid (11 %), plant sterols (0.53 - 0.97 %), and tocopherols (700 - 1,200 ppm) [2, 12, 13].

Rapeseed oil produced from wild seeds contains 50 % of potentially harmful for human health erucic acid and high levels of glucosinolates (mustard oil glycosides), that can be used as valuable and renewable raw material for production of a wide array of industrial products, as pesticides, anti-microbial, anti-fungicidal, anti-bacterial, and anti-virus agents for plant protection against pests and pathogens, and as biofumigants [11, 14 - 17].

Nowadays the plant breeders create the suitable for human nutrition and animal feed double low oilseed rape varieties containing decreased content of erucic acid in the rape oil to 0 % - 40 % and low level of glucosinolates, and vice versa increased content of linoleic acid to 15 - 20 % and linolenic acid to 8 % - 20 % [2, 9]. Recently, the European Food Safety

Authority allowed using rapeseed protein isolate as an important protein supplements to human foods [2, 18]. The oilseed rape oil glucosinolates can be used in medicine to prevent cancer due to their anti-cancer properties [2, 13, 18].

Oilseed rape oil-based derivatives are used for industrial purposes, including production of environmentally friendly and less toxic than traditional petroleum based derivatives: biofuels, printing inks, lubricants, stabilizers and processing aids, flame retardants in the manufacture of plastics, slip agents to prevent adhesion in polythene film, and anti-block, anti-static, plasticizing, and reactive agents used in the manufacture of polyamides and polyesters [2].

Oilseed rape meal proteins-based derivatives are also used as source for production of bioplastics, coatings, glue, adhesive, paper, cosmetic, emulsifier, as encapsulation agents for pharmaceuticals, agrochemicals and flavours, and as a combustion material. Rapeseed oil cake meal remaining after rapeseed oil production is enriched by high content of important amino acids such as methionine, cystine, and threonine, fatty acids, and minerals such as P, Ca, Mg, Mn and Se, therefore it is used as an animal feed and plant fertilizer [2, 7, 9-11, 18, 19].

Oilseed rape straw is used as animal bedding, animal feed, combustion, as a source for production of industrial fibre, and as a combustion material [2, 11].

Global climatic changes, soil contamination by industrial wastes and pesticides, pathogens and pests adversely affect the growth and decrease the yield of industrially important culture oilseed rape (*Brassica napus* L.) [5, 20, 21]. Today the natural and synthetic plant growth regulators, mineral fertilizers, and pesticides are widely used in agriculture sector of economically developed countries to accelerate oilseed rape growth and increase yield, and to protect from pests and adverse environmental factors [5, 22 – 31]. Nevertheless, the elaboration of new effective and ecologically friendly plant growth regulators on the base of natural and synthetic compounds to improve growth and development of this strategic energy and food crop is an actual problem for modern agriculture [32 - 34].

The most innovative approach to solve this actual problem is the elaboration of new plant growth regulators and plant protection agents on the basis of chemical low molecular weight heterocyclic compounds, derivatives of pyridine, pyrimidine, pyrazole, triazole, and oxazole [31, 35 – 48]. Today the new biologically active compounds are synthesized on the base of low molecular weight five and six-membered heterocyclic compounds in the Institute of Bioorganic Chemistry and Petrochemistry of NAS of Ukraine. Our previously conducted researchers showed the advisability of application of low molecular weight synthetic heterocyclic compounds, derivatives of pyridine, pyrimidine, pyrazolotriazine, oxazole and oxazolopyrimidine to accelerate growth of various important agricultural crops during their vegetation [49 – 55].

The present work is aimed to study the effect of new chemical low molecular weight heterocyclic compounds, derivatives of oxazole and oxazolopyrimidine on regulation of seed germination and growth of oilseed rape (*Brassica napus* L.) of cultivar Kalinivsky during plant vegetative stage.

## MATERIALS AND METHODS

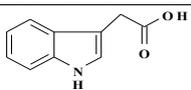
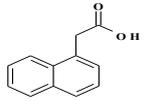
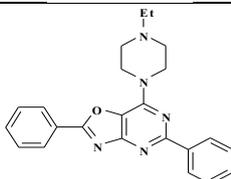
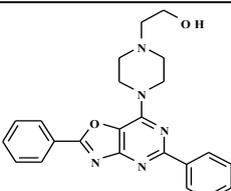
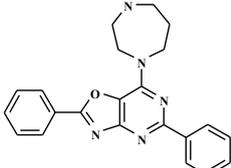
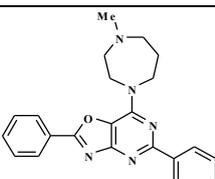
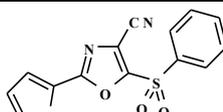
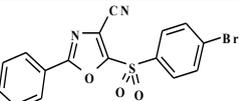
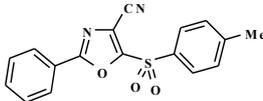
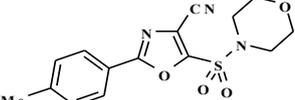
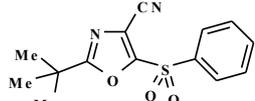
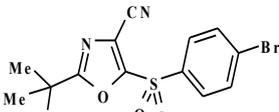
### Chemical structure of derivatives of oxazole and oxazolopyrimidine and phytohormones

In the laboratory conditions we studied the growth regulatory activity of chemical compounds, derivatives of oxazole and oxazolopyrimidine on intensification of germination of seeds and growth of oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky during the 21<sup>st</sup> day. These chemical compounds were synthesized at the Department for chemistry of bioactive nitrogen-containing heterocyclic compounds of Institute of Bioorganic Chemistry and Petrochemistry of NAS of Ukraine.

The growth regulatory activity of chemical compounds was compared with growth regulatory activity of plant hormones auxins IAA (1*H*-Indol-3-ylacetic acid) and NAA (1-Naphthylacetic acid).

The chemical structure, name and relative molecular weights of derivatives of oxazole and oxazolopyrimidine (the compounds № 1 - 10) and phytohormones auxins IAA and NAA are shown in Table 1.

**Table-1: The structure, name and relative molecular weight of tested chemical compounds**

№	Chemical structure of tested compound	Chemical name and relative molecular weight of tested compound
IAA		(1 <i>H</i> -Indol-3-ylacetic acid), MW=175.19
NAA		(1-Naphthylacetic acid), MW=186.21
1		7-(4-Ethylpiperazin-1-yl)-2,5-diphenyl[1,3]oxazolo[4,5- <i>d</i> ]pyrimidine, MW=385.47
2		2-[4-(2,5-Diphenyl[1,3]oxazolo[4,5- <i>d</i> ]pyrimidin-7-yl)piperazin-1-yl]ethanol, MW=401.47
3		7-(1,4-Diazepan-1-yl)-2,5-diphenyl [1,3]oxazolo[4,5- <i>d</i> ]pyrimidine, MW=371.45
4		7-(4-Methyl-1,4-diazepan-1-yl)-2,5-diphenyl[1,3]oxazolo[4,5- <i>d</i> ]pyrimidine, MW=385.47
5		5-(Phenylsulfonyl)-2-(2-thienyl)-1,3-oxazole-4-carbonitrile, MW=316.36
6		5-[(4-Bromophenyl)sulfonyl]-2-phenyl-1,3-oxazole-4-carbonitrile, MW=389.23
7		5-[(4-Methylphenyl)sulfonyl]-2-phenyl-1,3-oxazole-4-carbonitrile, MW=324.36
8		2-(4-Methylphenyl)-5-(morpholin-4-ylsulfonyl)-1,3-oxazole-4-carbonitrile, MW=333.37
9		2- <i>tert</i> -Butyl-5-(phenylsulfonyl)-1,3-oxazole-4-carbonitrile, MW=290.34
10		5-[(4-Bromophenyl)sulfonyl]-2- <i>tert</i> -butyl-1,3-oxazole-4-carbonitrile, MW=369.24

## Plant growing conditions and treatment

Seeds of oilseed rape (*Brassica napus* L.) of cultivar Kalinivsky were surface sterilized in 1 %  $\text{KMnO}_4$  solution for 3 min and 96 % ethanol solution for 1 min, and then washed three times with sterile distilled water. After this procedure seeds were placed in the cuvettes (each containing 25-30 seeds) on the perlite moistened with distilled water (control), or with water solution of derivatives of oxazole and oxazolopyrimidine, and phytohormones auxins IAA and NAA used at the same concentration  $10^{-9}\text{M}$ . After this procedure the control and experimental seeds were placed in the thermostat for their germination in the darkness at the temperature  $23\text{ }^\circ\text{C}$  during 48 hours. Sprouted seedlings were placed in the plant growth chamber in which seedlings were grown for 24 days at the 16/8 h light/dark conditions, at the temperature  $24\text{ }^\circ\text{C}$ , light intensity 3000 lux and air humidity 60-80 %. Comparative analysis of biometric indices of seedlings (i.e. number of germinated seeds (%), length of shoots (cm), total number of roots (pcs), total length of roots (mm)) was carried out at the 21<sup>st</sup> day after their sprouting according to the guideline [56].

The biometric indices determined in the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the solution of chemical compounds, derivatives of oxazole and oxazolopyrimidine used at the concentration  $10^{-9}\text{M}$  was expressed in % according to similar index determined in the control 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), or 21<sup>st</sup>-day-old oilseed rape seedlings grown on the solution of phytohormones auxins IAA and NAA used at the same concentration  $10^{-9}\text{M}$ .

## Determination of photosynthetic pigments content in the oilseed rape seedlings

The total content of chlorophyll a, chlorophyll b, and carotenoids was determined in the leaves of 21<sup>st</sup>-day-old oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky grown either on the distilled water (control), or on the water solution of chemical compounds, derivatives of oxazole and oxazolopyrimidine, and phytohormones auxins IAA and NAA used at the same concentration  $10^{-9}\text{M}$ .

The sample (500 mg) of leaves isolated from control and experimental 21<sup>st</sup>-day-old oilseed rape seedlings was homogenized in the porcelain mortar in a cooled at the temperature  $10\text{ }^\circ\text{C}$  96 % ethanol at the ratio of 1: 10 (weight: volume) with addition of 0,1-0,2 g  $\text{CaCO}_3$  (to neutralize the plant acids) to perform extraction of pigments. The 1 ml of homogenate was centrifuged at 8000 g in a refrigerated centrifuge K24D (MLW, Engelsdorf, Germany) during 5 min and at the temperature  $4\text{ }^\circ\text{C}$ . The obtained precipitate was washed three times with 1 ml 96 % ethanol and centrifuged at above mentioned conditions. After this procedure the optical density of chlorophyll a,

chlorophyll b and carotenoid in the obtained extract was measured using spectrophotometer Specord M-40 (Carl Zeiss, Germany). The total content of chlorophyll a, chlorophyll b, and carotenoids was calculated in accordance with formula [57]:

$$C_{\text{chl a}} = 13.36 \times A_{664.2} - 5.19 \times A_{648.6},$$

$$C_{\text{chl b}} = 27.43 \times A_{648.6} - 8.12 \times A_{664.2},$$

$$C_{\text{chl (a + b)}} = 5.24 \times A_{664.2} + 22.24 \times A_{648.6},$$

$$C_{\text{car}} = (1000 \times A_{470} - 2.13 \times C_{\text{chl a}} - 97.64 \times C_{\text{chl b}}) / 209,$$

Where,  $C_{\text{chl}}$  – concentration of chlorophylls (mg/ml),  $C_{\text{car}}$  – concentration of carotenoids (mg/ml),  $C_{\text{chl a}}$  – concentration of chlorophyll a (mg/ml),  $C_{\text{chl b}}$  – concentration of chlorophyll b (mg/ml),  $A$  – absorbance value at a proper wavelength in nm.

The chlorophyll content per 1 g of fresh weight (FW) of extracted from tomato leaves was calculated by the following formula (separately for chlorophyll a and chlorophyll b):

$$A_1 = (C \times V) / (1000 \times a_1),$$

Where,  $A_1$  – content of chlorophyll a or chlorophyll b (mg/g FW),  $C$  – concentration of pigments (mg/ml),  $V$  – volume of extract (ml),  $a_1$  – sample of plant tissue (g).

The index of content of chlorophyll a, chlorophyll b, and carotenoids determined in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the solution of derivatives of oxazole and oxazolopyrimidine used at the concentration  $10^{-9}\text{M}$  was expressed in % according to similar index determined in the leaves of control 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), or 21<sup>st</sup>-day-old oilseed rape seedlings grown on the solution of phytohormones auxins IAA and NAA used at the same concentration  $10^{-9}\text{M}$ .

## Statistical Analysis

Each experiment was performed in triplicate. Statistical analysis of the data was performed using dispersive Student's-t test with the level of significance at  $p \leq 0,05$ , the values are mean  $\pm$  Standard Deviation [58].

## RESULTS

### Impact of derivatives of oxazole and oxazolopyrimidine and phytohormones on growth of oilseed rape seedlings

As is known the major plant hormones auxins and cytokinins are involved in control of plant embryogenesis, seed germination, de-etiolation, cell cycle control, cell elongation and differentiation, protein synthesis, growth and development of plant root and shoot, development of flower and fruit, prevention of leaf abscission and delaying of leaf senescence [59 - 70].

In our work the regulatory effect of chemical compounds, derivatives of oxazole and oxazolopyrimidine, and phytohormones IAA and NAA used at the same concentration  $10^{-9}$ M on germination of seeds and growth of oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky during 21 days was studied.

The obtained results showed that all tested chemical compounds, derivatives of oxazole and oxazolopyrimidine revealed phytohormone-like regulatory effect on growth of roots and shoots on the 21<sup>st</sup>-day-old oilseed rape seedlings (Figure-1).



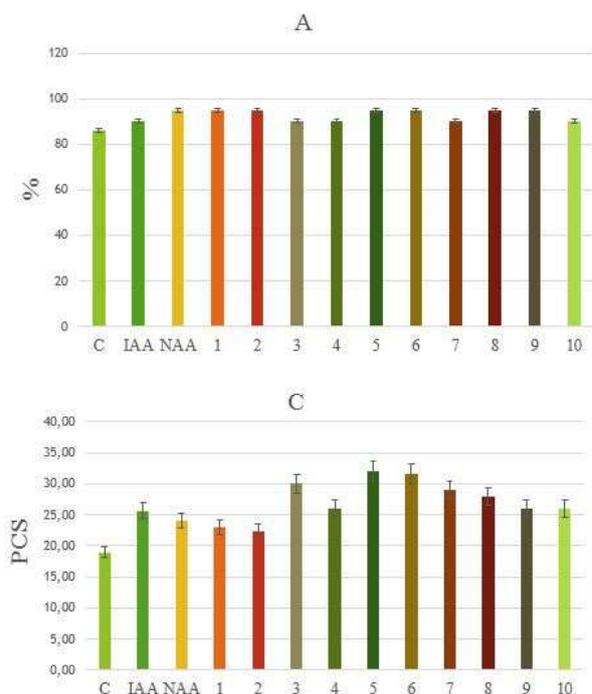
Figure-1 Effect of derivatives of oxazole and oxazolopyrimidine, and phytohormones IAA and NAA on growth of roots and shoots on the 21<sup>st</sup>-day-old oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky. C – Control (distilled water), the compounds № 1 – 10 - the derivatives of oxazole and oxazolopyrimidine, auxins IAA - 1*H*-Indol-3-ylacetic acid and NAA - 1-Naphthylacetic acid

The comparative statistical analysis of biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings (i.e. number of germinated seeds (%), length of shoots (cm), total number of roots (pcs), total length of roots (mm)) showed that the biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the water solution of chemical compounds, derivatives of oxazole and oxazolopyrimidine used at the concentration  $10^{-9}$ M were similar or higher than the biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown either on the distilled water (control), or on the water solution of auxins IAA (1*H*-Indol-3-ylacetic acid) and NAA (1-Naphthylacetic acid) used at the same concentration  $10^{-9}$ M (Figure 2).

It was found that biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of derivatives of oxazole and oxazolopyrimidine were increased by an average to 11 - 30 % – by length of shoots, by an average to 8 - 68 % – by total number of roots, and by an average to 5 - 43 % – by total length of roots, as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or on the  $10^{-9}$ M solution of plant hormones auxins IAA (1*H*-Indol-3-ylacetic acid) and NAA (1-Naphthylacetic acid) (Figure 2).

Among all tested chemical compounds, derivatives of oxazole and oxazolopyrimidine, some chemical compounds № 7, 9 and 10 revealed high growth regulatory activity on the indices of length of shoots which were increased by an average to 14 – 30 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), by an average to 11 - 28 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of plant hormone IAA (1*H*-Indol-3-ylacetic acid), and

by an average to 12 - 29 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape



seedlings grown on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure 2).

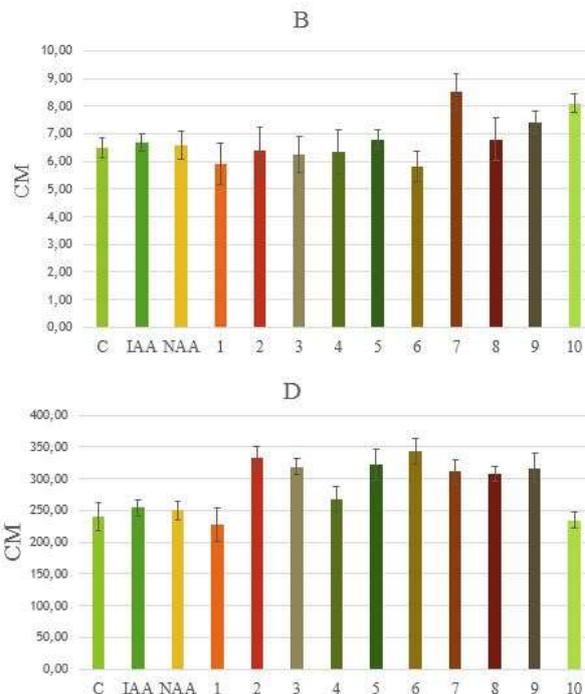


Figure-2 Effect of derivatives of oxazole and oxazolopyrimidine, and phytohormones IAA and NAA on biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky: A – number of germinated seeds (%); B – length of shoots (cm); C - total number of roots (psc); D - total length of roots (mm).

According to indices of total number of roots the highest growth regulatory activity among tested chemical compounds, derivatives of oxazole and oxazolopyrimidine revealed the compounds № 3, 5 and 6; the indices of total number of roots were increased by an average to 57 – 68 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), by an average to 17 – 25 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone IAA (1*H*-Indol-3-ylacetic acid), and by an average to 25 – 33 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure 2).

The lower growth regulatory activity according to indices of total number of roots revealed the compounds № 1, 2, 4, 7, 8, 9 and 10; the indices of total number of roots were increased by an average to 18 – 53 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), by an average to 9 – 13 % as compared with similar indices obtained on the 21<sup>st</sup>-day-

old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone IAA (1*H*-Indol-3-ylacetic acid), and by an average to 8 – 21 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure 2).

According to indices of total length of roots the highest growth regulatory activity among tested chemical compounds, derivatives of oxazole and oxazolopyrimidine revealed the compounds № 2, 3, 5 and 6; the indices of total length of roots were increased by an average to 33 – 43 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), by an average to 26 – 35 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone IAA (1*H*-Indol-3-ylacetic acid), and by an average to 28 – 38 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure-2).

The lower growth regulatory activity according to indices of total length of roots revealed the compounds № 1, 4, 7, 8 and 9; the indices of total length of roots were increased by an average to 11 – 32 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control), by an average to 5 – 25 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution

of plant hormone IAA (1*H*-Indol-3-ylacetic acid), and by an average to 7 – 27 % as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure-2).

The study of relationship between chemical structure and plant growth regulatory activity of oxazole and oxazolopyrimidine derivatives showed that biological activity of these compounds was differentiated depending on their chemical structure. The highest regulatory activity on growth of oilseed rape seedlings during 21 days was revealed for the compounds № 2 and 3 belonging to condensed derivatives of oxazole, and the compounds № 5 and 6 belonging to the uncondensed 5-arylsulfonyl-substituted oxazoles.

The comparative analysis of growth regulatory activity of compounds № 1 and 2 showed that the compound № 2 containing 1-ethanol-piperazine substituent in the 7 position of oxazolopyrimidine revealed higher regulatory activity on growth of oilseed rape seedlings during 21 days; the lower regulatory activity revealed the compound № 1 containing 4-ethylpiperazine substituent in the 7 position of oxazolopyrimidine fragment. Probably, the presence of hydroxyl substituent increases the activity of chemical compound № 2.

The comparative analysis of growth regulatory activity of compounds № 3 and 4 showed that the compound № 3, which does not contain methyl substituent in the diazepan fragment, revealed higher regulatory activity on growth of oilseed rape seedlings during 21 days; the lower regulatory activity revealed the compound № 4 containing methyl substituent in the diazepan fragment.

The comparative analysis of growth regulatory activity of compounds № 5 and 6 belonging to the uncondensed 5-arylsulfonyl-substituted oxazoles showed that the compound № 5 containing thiophene substituent in the 2 position of oxazole, revealed higher regulatory activity on growth of oilseed rape seedlings during 21 days; the lower regulatory activity revealed the compound № 6 containing phenyl substituent in the 2 position and 4-bromophenyl-sulfonyl substituent in the 5 position of oxazole.

The comparative analysis of growth regulatory activity of compounds № 6 and 7 showed the compound № 6 with bromine to be more active than compound № 7 without bromine.

The comparative analysis of growth regulatory activity of the compounds № 8, 9, and 10 belonging to uncondensed derivatives of oxazole, showed that the compound № 8 containing 4-

methylphenyl substituent in the 2 position and morpholin-4-yl-sulfonyl substituent in 5 position of oxazole, and compound № 9 with *tert*-butyl substituent in the 2 position and phenylsulfonyl substituent in the 5 position of oxazole revealed higher regulatory activity on growth of oilseed rape seedlings during 21 days; the lower regulatory activity revealed the compound № 10 containing 4-bromophenylsulfonyl substituent in the 5 position and *tert*-butyl substituent in the 2 position of oxazole.

Thus, the conducted researches showed that all tested chemical compounds, derivatives of oxazole and oxazolopyrimidine used at the concentration 10<sup>-9</sup>M revealed high growth regulating activity on germination of seeds and growth of oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky during 21 days. Obviously, that the high growth regulatory activity of derivatives of oxazole and oxazolopyrimidine is explained by their auxin-like and cytokinin-like stimulating effects on plant cell elongation, proliferation and differentiation, that are the basic processes of plant growth and development [59 - 70].

#### **Impact of derivatives of oxazole and oxazolopyrimidine and phytohormones on content of photosynthetic pigments in the leaves of oilseed rape seedlings**

As is known phytohormones cytokinins take an important part in regulation of biosynthesis in plant cells of the major photosynthetic pigments such as chlorophylls and carotenoids playing a key role in photosynthetic processes and photoprotection in plants, and providing plant productivity [57, 59, 61, 62, 68, 71-74].

In our experiments the phytohormone-like regulatory effects of chemical compounds, derivatives of oxazole and oxazolopyrimidine used at the concentration 10<sup>-9</sup>M, and phytohormones auxins IAA and NAA used at the same concentration 10<sup>-9</sup>M on synthesis of the major photosynthetic pigments (chlorophyll a, chlorophyll b, and carotenoids) in the 21<sup>st</sup>-day-old oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky were studied.

It was found that among all tested chemical compounds, derivatives of oxazole and oxazolopyrimidine, the highest regulatory activity revealed the chemical compounds № 4, 6, 8 and 9; the content of chlorophyll a and chlorophyll b in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of these compounds was increased by an average to 14 - 20 % – by content of chlorophyll a, by an average to 15 - 21 % – by content of chlorophyll b, by an average to 16 – 18 % – by content of chlorophyll a+b, as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) (Figure-3).

At the same time the content of chlorophyll a and chlorophyll b in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup>M solution of chemical compounds № 4, 6, 8 and 9 did not increase significantly as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of plant hormone IAA (1*H*-Indol-3-ylacetic acid) or on the 10<sup>-9</sup> M solution of plant hormone NAA (1-Naphthylacetic acid) (Figure 3).

It was also found that the content of carotenoids in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the 10<sup>-9</sup> M solution of chemical compounds № 4, 6, 8 and 9 was increased by an average to 14 - 26 % as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or grown on the 10<sup>-9</sup> M solution of IAA and NAA, respectively (Figure 3).

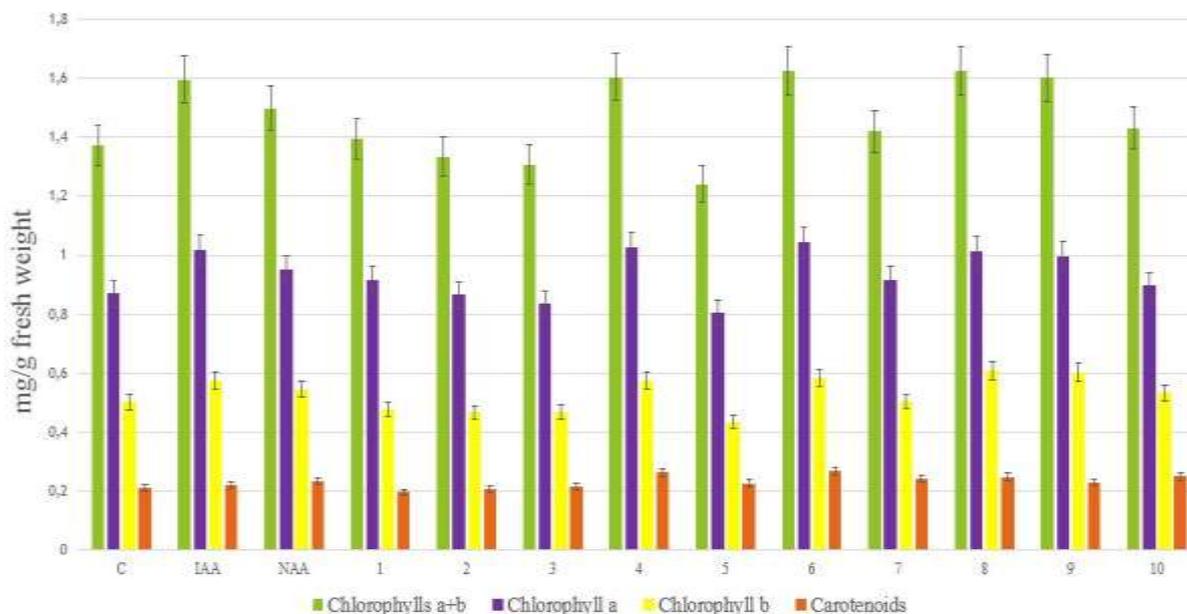


Figure-3 Effect of derivatives of oxazole and oxazolopyrimidine, and phytohormones IAA and NAA on content of chlorophyll a, chlorophyll b and carotenoids in the leaves of 21<sup>st</sup>-day-old oilseed rape (*Brassica napus* L.) seedlings of cultivar Kalinivsky

The study of relationship between chemical structure and regulatory activity of oxazole and oxazolopyrimidine derivatives on synthesis of photosynthetic pigments (chlorophylls a, b, and carotenoids) in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings showed that the highest regulatory activity was revealed for derivatives of oxazolopyrimidine – the compound № 4 containing methyl substituent in the diazepan fragment. At the same time the compound № 3 without methyl group in the diazepan fragment, did not exhibited regulatory activity.

Among the oxazole derivatives the high regulatory activity was revealed for compounds № 6, 8, and 9 with different substituents in the 2 and CN-group in 4 position of oxazole ring and similar arylsulfonyl or sulfonamide substituents in the 5 position of oxazole.

The comparative analysis of regulatory activity between non-condensed and condensed sulfonyl-substituted derivatives of oxazole showed that non-

condensed compounds exhibited higher regulatory activity than condensed derivatives of oxazole.

Thus the obtained results proved the positive effect of chemical compounds № 4, 6, 8 and 9, derivatives of oxazole and oxazolopyrimidine on increasing of content of photosynthetic pigments (chlorophylls a, b, and carotenoids) in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings playing an important role in photosynthetic processes and providing plant productivity [57, 71 - 74].

Possibly this fact is explained by cytokinin-like effect of pyrimidine derivatives on increasing of synthesis of chlorophylls and carotenoids and on delaying chlorophyll breakdown in in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings [59, 61, 62, 68, 71 - 74].

## CONCLUSION

Screening of new effective regulators among chemical compounds, derivatives of oxazole and oxazolopyrimidine to accelerate vegetative growth of oilseed rape (*Brassica napus* L.) of cultivar Kalinivsky was conducted. The growth regulatory activity of chemical compounds was compared with growth regulatory activity of plant hormones auxins IAA (1*H*-Indol-3-ylacetic acid) and NAA (1-Naphthylacetic

acid). The conducted researchers showed that all tested chemical compounds, derivatives of oxazole and oxazolopyrimidine used at the concentration  $10^{-9}$ M revealed the high stimulating effect on the growth of oilseed rape seedlings during 21 days. The biometric indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of derivatives of oxazole and oxazolopyrimidine were increased as compared with similar indices obtained on the 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or on the  $10^{-9}$  M solution of plant hormones auxins IAA and NAA. The study of relationship between chemical structure and plant growth regulatory activity of derivatives of oxazole and oxazolopyrimidine showed that growth regulatory activity of these compounds varied depending on their chemical structure. The highest growth regulatory activity on growth of oilseed rape seedlings during 21 days exhibited the derivatives of oxazolopyrimidine – the compounds № 2 and 3 belonging to condensed derivatives of oxazole, and the compounds № 5 and 6 belonging to the uncondensed 5-arylsulfonyl-substituted oxazoles. It was also found that compound № 4 – derivative of oxazolopyrimidine and compounds № 6, 8, and 9 – derivative of oxazole showed the high regulatory activity on increasing of synthesis of photosynthetic pigments (chlorophylls a, b, and carotenoids) in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings. The content of photosynthetic pigments in the leaves of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the  $10^{-9}$ M solution of chemical compounds № 4, 6, 8, and 9 was increased as compared with similar indices of 21<sup>st</sup>-day-old oilseed rape seedlings grown on the distilled water (control) or on the  $10^{-9}$ M solution of IAA and NAA, respectively. The obtained results proved the possibility of application of chemical compounds, derivatives of oxazole and oxazolopyrimidine as new effective regulators to improve the vegetative growth of oilseed rape (*Brassica napus* L.) of cultivar Kalinivsky.

#### Conflict of interest

Authors stated that there is no conflict of interest.

#### REFERENCES

1. Sovero, M. (1993). Rapeseed, a new oilseed crop for the United States. In: J. Janick and J.E. Simon (Eds.), *New crops*. Wiley, New York, 302-307.
2. Zahoor, F., Forristal, D., Teagasc, Park, O., Carlow. (2000). *Cropquest*. Oil seed rape: Crop report.
3. Zhang, M., & Malhi, S. S. (2010). Perspectives of oilseed rape as a bioenergy crop. *Biofuels*, 1(4), 621-630.
4. Moser, D., Eckerstorfer, M., Pascher, K., Essl, F., & Zulka, K. P. (2013). Potential of genetically modified oilseed rape for biofuels in Austria: Land use patterns and coexistence constraints could decrease domestic feedstock production. *Biomass and bioenergy*, 50, 35-44.
5. Berry, P., Cook, S., Ellis, S., Gladders, P., & Roques, S. (2014). *HGCA Oilseed rape guide*. E. Boys (Ed.). HGCA Publications.
6. van Duren, I., Voinov, A., Arodudu, O., & Firrisa, M. T. (2015). Where to produce rapeseed biodiesel and why? Mapping European rapeseed energy efficiency. *Renewable energy*, 74, 49-59.
7. Balalić, I., Marjanović-Jeromela, A., Crnobarac, J., Terzić, S., Radić, V., Miklič, V., & Jovičić, D. (2017). Variability of oil and protein content in rapeseed cultivars affected by seeding date. *Emirates Journal of Food and Agriculture*, 29(6), 404.
8. Riffkin, P. A., O'Leary, G., & Acuña T. (2012). The effects of plant growth regulators on winter and spring canola (*Brassica napus* L.) types in the High Rainfall Zone of south-eastern Australia. Proceedings of 16th Australian Agronomy Conference "Capturing opportunities and overcoming obstacles in Australian agronomy". 14-18 October 2012, Armidale, New South Wales, Australia, 67.
9. Bagger, C.L., Bellostas, N., Jensen, S.K., Sørensen, H., Sørensen, J.C., Sørensen, S. (2007). Processing-bioprocessing of oilseed rape in bioenergy production and value added utilization of remaining seed components. Feed and industrial raw material: Industrial Materials and Biofuel. Proceedings of the 12th International Rapeseed Congress. Volume V. Sustainable Development in Cruciferous Oilseed Crops Production. Wuhan, China. March 26-30, 2007, Science Press USA Inc., 315.
10. Rossato, L., Laine, P., & Ourry, A. (2001). Nitrogen storage and remobilization in *Brassica napus* L. during the growth cycle: nitrogen fluxes within the plant and changes in soluble protein patterns. *Journal of Experimental Botany*, 52(361), 1655-1663.
11. Weightman, R. (2014). *Nutritional Value of Oilseed Rape and Its Co-products for Pig and Poultry: Potential Improvements and Implications for Plant Breeders*. HGCA.
12. Jahreis, G., Schäfer, U. (2011). Rapeseed (*Brassica napus*) Oil and its Benefits for Human Health. P. 967-974. In: *Nuts and Seeds in Health and Disease Prevention*. V.R. Preedy, R.R. Watson, V.B. Patel (Eds.). Elsevier Inc., 477.
13. Lin, L., Allemekinders, H., Dansby, A., Campbell, L., Durance-Tod, S., Berger, A., & Jones, P. J. (2013). Evidence of health benefits of canola oil. *Nutrition reviews*, 71(6), 370-385.
14. Bhardwaj, H. L., & Hamama, A. A. (2000). Oil, erucic acid, and glucosinolate contents in winter hardy rapeseed germplasms. *Industrial Crops and Products*, 12(1), 33-38.

15. Zukalová, H., & Vasak, J. (2002). The role and effects of glucosinolates of Brassica species—a review. *Rostlinna Vyroba*, 48(4), 175-180.
16. Bhardwaj, H. L., & Hamama, A. A. (2003). Accumulation of glucosinolate, oil, and erucic acid in developing Brassica seeds. *Industrial crops and products*, 17(1), 47-51.
17. Špak, J., Kolářová, L., Lewis, J., & Fenwick, G. R. (1993). The effect of glucosinolates (mustard oil glycosides) and products of their enzymic degradation on the infectivity of turnip mosaic virus. *Biologia plantarum*, 35(1), 73.
18. Nosenko, T., Kot, T., & Kichshenko, V. (2014). Rape seeds as a source of feed and food proteins. *Polish journal of food and nutrition sciences*, 64(2), 109-114.
19. Kracht, W., Dänicke, S., Kluge, H., Keller, K., Matzke, W., Hennig, U., & Schumann, W. (2004). Effect of dehulling of rapeseed on feed value and nutrient digestibility of rape products in pigs. *Archives of animal nutrition*, 58(5), 389-404.
20. Diepenbrock, W. (2000). Yield analysis of winter oilseed rape (*Brassica napus* L.): a review. *Field Crops Research*, 67(1), 35-49.
21. Berry, P. M., & Spink, J. H. (2006). A physiological analysis of oilseed rape yields: past and future. *The Journal of Agricultural Science*, 144(5), 381-392.
22. Armstrong, E. L., & Nicol, H. I. (1991). Reducing height and lodging in rapeseed with growth regulators. *Australian Journal of Experimental Agriculture*, 31(2), 245-250.
23. Child, R. D., Evans, D. E., Allen, J., & Arnold, G. M. (1993). Growth responses in oilseed rape (*Brassica napus* L.) to combined applications of the triazole chemicals triapenthenol and tebuconazole and interactions with gibberellin. *Plant growth regulation*, 13(2), 203-212.
24. Spink, J. H., Rosemaund, A., & Stokes, D. T. (1997). Evaluation of the effect of plant growth regulators and fungicides on light interception, growth and yield of oilseed rape. HGCA Project report OS24, 14.
25. Kirkland, K. J. (1992). Effect of triapenthenol plant-growth regulator on canola height, yield, side branches and pod density. *Canadian Journal of Plant Science*, 72(4), 1153-1156.
26. Gaveliene, V., Novickiene, L., Miliuviene, L., Brazauskiene, I., & Kazlauskiene, D. (2005). Possibilities to use growth regulators in winter oilseed rape growing technology 2. Effects of auxin analogues on the formation of oilseed rape generative organs and plant winterhardiness. *Agronomy research*, 3(1), 9-19.
27. Pits, N., Kubacki, K., & Tys, J. (2008). Influence of application of plant growth regulators and desiccants on a yield and quality of winter oilseed rape. *International Agrophysics*, 22(1), 67.
28. Matysiak, K., & Kaczmarek, S. (2013). Effect of chlorocholine chloride and triazoles—tebuconazole and flusilazole on winter oilseed rape (*Brassica napus* var. *oleifera* L.) in response to the application term and sowing density. *Journal of plant protection research*, 53(1), 79-88.
29. Ijaz, M., Mahmood, K., & Honermeier, B. (2015). Interactive Role of Fungicides and Plant Growth Regulator (Trinexapac) on Seed Yield and Oil Quality of Winter Rapeseed. *Agronomy*, 5(3), 435-446.
30. Jankowski, K. J., Sokólski, M., Dubis, B., Krzebietke, S., Żarczyński, P., Hulanicki, P., & Hulanicki, P. S. (2016). Yield and quality of winter oilseed rape (*Brassica napus* L.) seeds in response to foliar application of boron. *Agricultural and Food Science*, 25(3), 164-176.
31. Berry, P. M., & Spink, J. H. (2009). Understanding the effect of a triazole with anti-gibberellin activity on the growth and yield of oilseed rape (*Brassica napus*). *The Journal of Agricultural Science*, 147(3), 273-285.
32. Basra, A.S. (Ed.). (2000). *Plant Growth Regulators in Agriculture and Horticulture: Their Role and Commercial Uses*. Haworth Press, Inc., 264.
33. Rademacher, W. (2015). Plant growth regulators: backgrounds and uses in plant production. *Journal of plant growth regulation*, 34(4), 845-872.
34. Lopez-Lauri, F. (2016). *Plant Growth Regulators*. In: Siddiqui, M.W., Zavala, A., Hwang, J.F., Andy, C.A. (Eds.). *Postharvest Management Approaches for Maintaining Quality of Fresh Produce*, Springer, 125-139.
35. Taylor, E. C., Wipf, P. (Eds.). *Chemistry of Heterocyclic Compounds: A series of monographs*. Series Online ISSN: 1935-4665.
36. Scriven, E. F., & Murugan, R. (2005). *Pyridine and pyridine derivatives*. *Kirk-Othmer Encyclopedia of Chemical Technology*.
37. Joule, J. A., & Mills, K. (2012). *Heterocyclic chemistry at a glance*. John Wiley & Sons.
38. Armstrong, S., Barnes, N. J., Barnett, S. P., Clarke, E. D., & Crowley, P. J. (2001). Isothiazole derivatives and their use as pesticides. Patent WO 2001055144 A1.
39. Baum, J.S., Chen, T.M. (1987). Plant growth and development modification using benzoxazole derivatives. Patent US 4659360 A.
40. Cansev, A., Gulen, H., Zengin, M.K., Ergin, S., Cansev, M., Kumral, N.A. (2016). Use of pyrimidines in stimulation of plant growth and development and enhancement of stress tolerance. Patent US 20160000075.
41. Corsi C., Wendeborn S.V., Bobbio C., Kessabi J., Schneiter P., Grasso V., Haas U.J. (2011). Isothiazole and pyrazole derivatives for use as plant growth regulators. Patent EP 2358699A1.
42. Dai, H., Li, Y. Q., Du, D., Qin, X., Zhang, X., Yu, H. B., & Fang, J. X. (2008). Synthesis and biological activities of novel pyrazole oxime derivatives containing a 2-chloro-5-thiazolyl

- moiety. *Journal of agricultural and food chemistry*, 56(22), 10805-10810.
43. Kuragano, T., & Tanaka, Y. (2002). Dérivés de la pyrimidine et herbicides les contenant. Patent WO 2002038550 A1.
44. Minn, K., Dietrich, H., Dittgen, J., Feucht, D., Häuser-Hahn, I., & Rosinger, C. H. (2008). Pyrimidine derivatives and their use for controlling undesired plant growth. Patent US 8329717 B2.
45. Newton, T., Waldeck, I. (2000). Oxazole carboxamide herbicides. Patent US6096688 A.
46. Nimbalkar, S., & Hote, S. V. (2015). Pyrazole Derivatives and their Synthesis-A review. *International Journal on Recent and Innovation Trends in Computing and Communication*, 3(2), 61-65.
47. Whittingham, W. G., Winn, C. L., Glithro, H., Boussemghoune, M. A., & Aspinall, M. B. (2010). Pyrimidine derivatives and their use as herbicides. WO Patent 2010092339 A1.
48. Zhao, Q., Liu, S., Li, Y., & Wang, Q. (2009). Design, synthesis, and biological activities of novel 2-cyanoacrylates containing oxazole, oxadiazole, or quinoline moieties. *Journal of agricultural and food chemistry*, 57(7), 2849-2855.
49. Tsygankova, V., Andrusevich, Y., Shtompel, O., Myroljubov, O., Hurenko, A., Solomyanny, R., ... & Brovarets, V. (2016). Study of Auxin, Cytokinin and Gibberellin-like Activity of Heterocyclic Compounds Derivatives of Pyrimidine, Pyridine, Pyrazole and Isoflavones. *European Journal of Biotechnology and Bioscience*, 4(12), 29-44.
50. Tsygankova, V., Andrusevich, Y., Shtompel, O., Hurenko, A., Solomyannyj, R., Mrug, G., ... & Brovarets, V. (2016). Stimulating effect of five and six-membered heterocyclic compounds on seed germination and vegetative growth of maize (*Zea mays* L.). *International Journal of Biology Research*, 1(4), 1-14.
51. Tsygankova, V., Andrusevich, Ya., Shtompel, O., Pilyo, S., Prokopenko, V., Kornienko, A., Brovarets V. (2016). Study of growth regulating activity derivatives of [1,3]oxazolo[5,4-d]pyrimidine and N-sulfonyl substituted of 1,3-oxazoles on soybean, wheat, flax and pumpkin plants. *International Journal of Chemical Studies*, 4(5), 106-120.
52. Tsygankova, V. A., Bayer, O. O., Andrusevich Ya, V., Galkin, A. P., Brovarets, V. S., Yemets, A. I., & Blume Ya, B. (2016). Screening of five and six-membered nitrogen-containing heterocyclic compounds as new effective stimulants of *Linum usitatissimum* L. organogenesis in vitro. *Int J Med Biotechnol Genetics* S, 2, 1-9.
53. Tsygankova, V., Andrusevich, Ya., Shtompel, O., Romaniuk, O., Yaikova, M., ...and Brovarets, V. (2017). Application of Synthetic Low Molecular Weight Heterocyclic Compounds Derivatives of Pyrimidine, Pyrazole and Oxazole in Agricultural Biotechnology as a New Plant Growth Regulating Substances. *Int J Med Biotechnol Genetics*, S2(002), 10 – 32.
54. Tsygankova, V.A., Andrusevich, Ya.V., Shtompel, O.I., Kopich, V.M., Pilyo, S.G., Prokopenko, V.M, Kornienko, A.M, Brovarets, V.S. (2017). Intensification of Vegetative Growth of Cucumber by Derivatives of [1,3]oxazolo[5,4-d]pyrimidine and N-sulfonyl substituted of 1,3-oxazole. *Research Journal of Life Sciences, Bioinformatics, Pharmaceutical, and Chemical Sciences (RJBPCS)*, 3(4), 107–122.
55. Tsygankova, V., Andrusevich, Y. A., Shtompel, O., Kopich, V., Solomyanny, R., Bondarenko, O., & Brovarets, V. (2018). Phytohormone-like effect of pyrimidine derivatives on regulation of vegetative growth of tomato. *International Journal of Botany Studies*, 3(2), 91-102.
56. Voytsehovska, O. V., Kapustyan, A. V., Kosik, O. I., Musienko, M. M., Olkhovich, O. P., & Panyuta, O. O. (2010). *Plant Physiology: Praktykum*, Teren, Lutsk, Ukraine, 420.
57. Lichtenthaler, H. K. (1987). Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Methods in Enzymology*, 148, 350-382.
58. Bang, H., Zhou, X. K., van Epps, H. L., & Mazumdar, M. (2010). *Statistical Methods in Molecular Biology*. Series: Methods in molecular biology. Humana press, New York, USA, 636.
59. Mok, M. C. (1994). Cytokinins and Plant Development – An Overview. *Cytokinins Chemistry, Activity, and Function*, 155-165.
60. Hoad G.V. Lenton J.R. Jackson M.B. (1987). *Hormone Action in Plant Development — A Critical Appraisal*. 1<sup>st</sup> Edn. Butterworth-Heinemann, 334.
61. Mok, D. W. S., & Mok, M. C. (2001). Cytokinin metabolism and action. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 52, 89-118.
62. Haberer, G., & Kieber, J. J. (2002). Cytokinins. New insights into a classic phytohormone. *Plant Physiology*, 128(2), 354-362.
63. Tsygankova, V. A., Galkina, L. A., Musatenko, L. I., & Sytnik, K. M. (2005). Genetic and epigenetic control of plant growth and development. Genes of auxin biosynthesis and auxin-regulated genes controlling plant cell division and extension. *Biopolymers and Cell*, 21(2), 107-133.
64. Hedden, P., & Thomas, S. G. (Eds.). (2006). *Plant Hormone Signaling*. Blackwell Publishing Ltd., 339.
65. Zhao, Y. (2010). Auxin biosynthesis and its role in plant development. *Annual review of plant biology*, 61, 49-64.
66. Sauer, M., Robert, S., & Kleine-Vehn, J. (2013). Auxin: simply complicated. *Journal of experimental botany*, 64(9), 2565-2577.
67. Wang, Y. H., & Irving, H. R. (2011). Developing a model of plant hormone interactions. *Plant signaling & behavior*, 6(4), 494-500.

68. Zwack, P. J., & Rashotte, A. M. (2013). Cytokinin inhibition of leaf senescence. *Plant signaling & behavior*, 8(7), e24737.
69. Lam-Son, T., Sikander, P. (Eds.). (2014). *Phytohormones: A Window to Metabolism, Signaling and Biotechnological Applications*. Springer-Verlag, 361.
70. Gao, Y. A., & Zhao, Y. U. (2014). Chapter 2 Auxin Biosynthesis and Catabolism. In: *Auxin and Its Role in Plant Development*. Zazimalova E., Petrasek J., Benkova E.(Eds.), Springer-Verlag Wien, VII: 444.
71. Bartley, G. E., & Scolnik, P. A. (1995). Plant carotenoids: pigments for photoprotection, visual attraction, and human health. *The Plant Cell*, 7(7), 1027.
72. Cazzonelli, C. I. (2011). Carotenoids in nature: insights from plants and beyond. *Functional Plant Biology*, 38, 833–847.
73. Gratani, L. (2014). Plant Phenotypic Plasticity in Response to Environmental Factors. *Advances in Botany*, Article ID 208747, 1-17.
74. Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D., & Darnell, J. (2000). *Molecular Cell Biology*. Section 16.3, Photosynthetic Stages and Light-Absorbing Pigments, 4<sup>th</sup> Ed., W.H. Freeman and Company, New York, USA.