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Ecotoxicological Evaluation of Commercial Fish Farm in Ogbogoro, Rivers State Nigeria: Gill Histological Assessment of *Clarias gariepinus* (African Catfish) Paul Chikwuogwo Wokpeogu, *Allison, Theodore Athanasius

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a concurrent increase in commercial fish farm. Nonetheless, there seems to be inadequate or non-regulation in the activities of commercial fish farming for the protection of public health. Hence the wholesomeness of fish consumed from commercial fish farms in Nigeria is highly questionable.

Fish are truly products of their environment, unlike land dwelling animals, fish metabolism and biological functions are directly linked to the physical and chemical properties of the medium that surrounds them; water [1].

It was observed, according to Olaleye *et al.*, [2], that of over 30,000 MT of various freshwater and brackish water fish species caught in the year 2000, catfishes were more abundant next to tilapias record revealed that the 46,206MT of catfishes were produced in the year 2007. These were consumed locally. With the present population of over 170 million, a projected

increase at an annual growth rate of 3.2% and the expected increase in fish demand.

Study area

Experimental Area (Ogbogoro commercial fish pond). Ogbogoro is a community, located in Obior Akpor Local Government Area of Rivers state, Nigeria. It is bounded by, Choba, Rumuekini, Emohua and Diobu. The geographical coordinates are, 4° 50¹ 48¹¹North, 6° 55¹ 50¹¹ East in DMS- Degrees Minutes Seconds or 4.8451°N, 6.9290°E, [in decimal degrees] (maplandia.com, 2017).



Fig-1: A map showing Ogbogoro community (adopted from maplandia.com)

Study area environment: the pond is sited in a swampy area which is prone to flood during the midraining season. It is located far from residential building. Aside the fishing section in the establishment, there are sections for piggery and poultry farming, although no animals were in them as of the time of my harvest of fishes for my experiment.

The pond is divided into units according to fig. 1.2. Each of this units has nothing less than 200 fishes in them. The fishes in each unit vary in size/age ranging from, hatchling, fingerling, nursery, table size.

Water regulation: According to koi food, the ideal regime for water changes is 10% per week or 20% per two weeks or 50% every six weeks. The percentage of water changed depends on the level of haze or cloudy appearance seen on the water, which is sign of high organics, (aquamed.com, 2000). 75% - 80% water must be changed at least twice per year. In ogbogoro, the pond water is changed every two weeks, completely, although varies depending on the level of pollution of the water.



Fig-2: Stagnant Concrete Pond in Ogbogoro (study site)

Feeding

Commercial fish feeds are concentrated, and if too much is eaten, it will pass through the fish only partly digested and then pollute the water. If excess food goes uneaten, it should be netted out to avoid pollution and feed less the next time. Ogbogoro commercial fish pond, feed their fishes during the day with fish feeds like coppens.

In terms of culture, because of the cannibalistic nature of Clarias gariepinus, multiple sorting is essential. As the fish grow, big ones of the same sizegroup are removed to another tank for rearing. Thus harvesting is done at different periods for the different groups sorted.

Reference Area (ARAC)

African Regional Aquaculture Center (ARAC), was chosen as the control site. It is situated at the training center, Omuihuechi, Aluu in Ikwere Local Government Area of Rivers State. Most of the activities in the center include research, training, and development of sustainable aquaculture options, in sub Saharan African. It covers an area of 81 hectares of land.

It's a centre of excellence that focuses on multidisciplinary approach to user-driven aqua cultural research, development and training in sub-Saharan Africa geared towards sustainable fish production in the region.

ARAC is affiliated to Rivers State University [RSU] for the award of masters of Science [M.Sc.] and post graduate diploma [PGD] in aquaculture.



Fig- 3: Earthen pond; Control site (ARAC)



Fig-4: Clarias gariepinus

African catfish (Clarias gariepinus) is one of the most important primary treatment for tropical cultured fish due to high growth rate, high stockingdensity capacities, and high resistance to poor water quality and oxygen and considered as a model for Eco toxicological studies [3].

Natural Distribution

They are found throughout Africa and the Middle East, and live in freshwater lakes, rivers, and swamps, as well as human-made habitats, such as oxidation ponds or even urban sewage systems. The African sharp tooth catfish was introduced all over the world in the early 1980s for aquaculture purposes, so is found in countries far outside its natural habitat, such as Brazil, Vietnam, Indonesia, and India.

Habitat

It is a nocturnal fish like many catfish. It feeds on living, as well as dead, animal matter. Because of its wide mouth, it is able to swallow relatively large prey whole. It has been known to take large water birds such as the common moorhen. It is also able to crawl on dry ground to escape drying pools. Further, it is able to survive in shallow mud for long periods of time, between rainy seasons. African catfish sometimes produce loud croaking sounds, not unlike the voice of the crow[4].

Natural spawning

Spawning mostly takes place at night in the shallow, inundated areas of the rivers, lakes and streams. Courtship is preceded by highly aggressive encounters between males. Courtship and mating takes place in shallow waters between isolated pairs of males and females. The male lies in a U-shape curved around the head of the female, held for several seconds. A batch of malt and eggs is released followed by a vigorous swish of the female's tail to distribute the eggs over a wide area. The pair usually rests after mating (from seconds up to several minutes) and then resumes mating [5].

There have been works on ecotoxicology which Contamination, pollutants, bio-markers and stressors were the focus [5-8, 5,9, 1, 10-13, 4, 9, 14-17].

Statement of the problem

High proliferation of fish farm business in Nigeria with inadequate knowledge of sources and breeding pattern and Inadequate or non-regulation of the fish farm business in Nigeria for protection of public health.

Aim

This study was aimed at determining the impact of the commercial fish pond in Ogbogoro, on the health of the cultivated fish.

Objectives

To determine qualitative histological analysis of target organ, the semi-quantitative histological analysis of target organs and the pollution status of the fish pond.

MATERIALS AND METHODS

Phase One (Preliminary Study)

The experimental site was inspected and vital questions were asked as: the number of fishes contained in the pond, type and frequency of fish feed used, treatment administered to fish in poor health condition, mode and frequency of changing the water content of the pond.

A sample fish was harvested and taken to the African Regional Aquaculture Centre for identification by a taxonomist.

Phase Two (Fish Sampling)

Fish Sampling

According to Institute of Veterinary Research and Food Security, Tirana, Albania, The European standards for fish sampling in lakes determined the sampling protocols and methodology developed in the course of fish and fishery monitoring for Prespa lakes. The sampling procedure was based on stratified random sampling.

Control

Control fishes were harvested. This was done by first collecting some water content of the pond into a plastic container which would contain the fishes from the control site to the laboratory. The essence was so that the original aquatic habitat of the fishes will remain the same after harvesting as it was before. Failure to do this will lead to alteration of the fish habitat and questionability of the results which will be gotten. Next, the remaining water content of the pond was drained, then with the aid of a seine; ten table-sized cat fishes were harvested and put into the plastic container in which there was exactly the same water content of the pond.

Study Site

Experimental fishes were harvested. This was done by first getting a good quantity of water from the

pond into a well-aerated plastic container designed for the purpose of transporting the fishes to the laboratory. The water content of the pond was drained in order to make the fishes more accessible for a good harvest. Twenty table-sized cat fishes were harvested from the pond

Harvesting of catfish

According to the United Nation's Food and Agricultural Organization, the following are the steps involve in the harvesting of catfish.

water in the pond was drained to concentrate the catfish, the right catfish seine was chosen and loaded to a seine reel, immersed in the deep end of the pond, and the seine was drawn the pond's harvesting area. The harvest was done at random and seining stopped when the catfish seine was full. The process was repeated till the required sample of size was caught. The sample size was confirmed by measuring on a weighing scale and transported to the laboratory.

Preparation of the Sample

According to American Veterinary Medical Association (AVMA), an acceptable method of euthanasia renders an animal unconscious and insensitive to pain and distress as quickly as possible, followed by cessation of all respiratory and circulatory functions and brain activity.

In line with AVMA guidelines the fishes were sacrificed through cervical dislocation method (severing the spinal cord anterior to the dorsal fin). Then the fishes were surgically opened on the ventral side of the fish for excision of the organs.

Histological Assessment

About 20 table-sized fish were harvested from Ogbogoro commercial fish pond and 10 table sized fish were also harvested from ARAC fish pond and organs of interest (gill, liver, and kidney) were extracted for histological assessment. This assessment involves the microscopic study of the tissues and is divided into two qualitative and semi quantitative analysis.

Qualitative Analysis

Tissue Processing

This assessment involves histological processes in order to view the cells of harvested organs with the aid of a light microscope, these processes are as follows:

STEP 1 (Pithing)

This is done by positioning the pointer end of a knife above its brain. Push it down quickly into the brain cavity of the fish. Locate the spot marked on the head, cut through with a sharp knife.

STEP 2 (Dissection)

Cut a slit along the belly of the catfish all the way to the anal fin. This is done with the aid of a dissecting kit (i.e., a scalpel and a scissors to be precise) to harvest the organs of interest from the sample fish while the heart of the fish is still beating.

Step 3 (Fixation)

This is done immediately after the organ of interest has been harvested from the sample fish, it involves transferring the harvested organ into a specimen bottle containing a fixative in this case 10% formal saline was used as the fixative. Fixatives aid in the preserving the tissue architecture of the organ and prevent putrefaction.

Step 4 (Dehydration)

This is the removal of water molecule from the fixed tissue with graded alcohol and aid the penetration of the clearing agent since it is not miscible with water. Usually harvested organ is passed through 50% alcohol, then 70%, 90%, and 100% (absolute) so as to remove the water molecule gradually and steadily but in this study the fish organs were place in 70%, 90%, 95% and 100% for 6 hours in order for the organs to dehydrate well, this is due to the fact since they are fishes they live in water there will be large amount of water quantity in their organs.

Step 5 (Clearing)

Here, a clearing agent in this case xylene is used to remove the alcohol in order to aid the penetration of paraffin wax since paraffin wax is immiscible with alcohol. The dehydrated tissue is the placed in a glass specimen bottle containing the clearing agent (xylene).

Step 6 (Impregnation/Embedding)

This involves placing the tissue cassette and place in melted wax in order for the paraffin wax to replace the water molecules and make the tissue hard for sectioning after which the tissue is placed in mould containing molten wax and allowed to solidify in the paraffin wax thereby forming a tissue block. The tissue block is then cast to a wooden lock in order to make it easy to be placed in a microtome for sectioning.

Step 7 (Sectioning)

This used to produce very thin slice (sections) of the tissue block in ribbons (which would be viewed under a light microscope) with the aid of a microtome. Sectioning makes it easy for tissue to be stained.

Step 8 (Staining)

This involves rinsing the thin tissue section to water through xylene, graded alcohol and then distilled water (i.e. 100%, 90%, 70%, and 50%) and then the section is stained with the H & E stain which is used to view the tissue component.

STEP 9 (Coverslipping)

This involves using a cover slip to cover the tissue section on the slide, and then it is mounted for viewing with the aid of a light microscope.

Semi Quantitative Histological Analysis

A qualitative assessment protocol was used to quantify histopathological alterations observed in the sections of each of the organ. A qualitative histopathological assessment was done using CX1 Olympus light microscope. Tissue sections were scanned on 400x magnification. The result was semiquantitatively assessed using part of a scoring system. Bernet *et al.* [19] modified from the protocol by [18]. In brief, the tissue samples were assessed by identifying histopathological alteration in terms of reaction patterns including:

- Circulatory disturbance
- Regressive changes
- Progressive changes
- Inflammatory responses
- Neoplasia; if identified, the alteration was given an importance factor which represents the potential of the alteration to affect fish health: 1 (alteration is reversible; 2 (alteration is reversible if the stressor is neutralised); 3 (alteration is irreversible). A score value, representing the occurrence of the alteration throughout the tissue was also assigned: 0 (absent), 2 (mild), 4 (moderate), and 6 (severe). The score value and the importance factor for each alteration were multiplied and these results for all the alterations identified in a single organ were then summed to give an organ index per fish.

Mathematical calculations of lesion indices:

Where org= organ; rp= reaction pattern (constant); alt= alterations; a= score value; w = importance factor.

• Organ index: The organ index (Iorg) represents the extent of damage to an organ. It allows for comparison of the extent of damage of the same organ in different individuals and is calculated as follows:

Iorg= $\sum T \sum alt$ (aorg rp alt x worg rp alt).

• Total fish index:

The fish index (Ifish) signifies a measure of the overall health status based on the lesions observed. It is also possible to compare individuals as the Ifish for each fish is calculated the same way:

If $ish = \Sigma \operatorname{org} \Sigma \operatorname{rp} \Sigma \operatorname{alt}$ (a or g r p alt x wor g r p alt).

Furthermore, a modified classification system by [20] based on a scoring scheme by Zimmerli *et al.* [21] was employed to evaluate the degree of histological changes. This classification system is based on the calculated mean organ index values.

Class 1 (index value <10): Slight histological alterations.

Class 2 (index value 10-25): Moderate histological alterations.

Class 3 (index value 26-35): Pronounced alterations of organ tissue. Class 4 (index value >35): Severe alterations of organ tissue.

STATISTICAL ANALYSIS

One way anova statistical method was used to analyse the organ indices of the specimens from Ogbogoro and ARAC at a significant level of 0.005

RESULTS AND DISSCUSION Results

Table-1: The percentage prevalence of Gills histopathology of fishes harvested from Ogbogoro and ARAC

Alteration	Prevalence (%)		
	OGBOGORO	ARAC	
	(n=20)	(n=10)	
Circulatory Disturbance (CD)			
Hyperaemia	13.75	7.5	
Haemorrhage	11.25	16.4	
Vacuolation	23.75	28.4	
Regressive Change (RC)			
Structural alterations	32.5	38.8	
Necrosis	7.5	0	
Progressive Change (PC)			
Epithelial Lifting	11.25	8.96	
AVERAGE % PREVALENCE	16.67	16.68	

Percentage prevalence for each alteration is calculated by the total number of each alteration divided

by the overall sum of the organ alterations, multiplied by 100.

Fable-2	: Statistical	analysis of	f organ in	dex (using	One Wa	ay Anova)
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Gill	Ν	Mean	Standard Deviation	F-test	Significance
1.00	10	16.000	3.771		
2.00	20	18.400	8.171	0.770	0.388
Total	30	17.600	7.034		
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The analysis showed that there is no significant difference between the two data (P>0.05)

Qualitative histological assessment results showed that circulatory disturbances (CD) and regressive changes (RC) were identified in gill tissue. Figure 5 represents a microscopic photograph of the gill tissue for *C. gariepinus*. CD included hyperemia, haemorrhage, vacuolation and epithelial lifting while RC included architectural and structural alterations of epithelial cells were so high with samples from experimental site (Figure 5: b-e) and necrosis. Structural changes in the form of fusion of secondary lamellae and fusion of adjacent lamella were also noted (Figure 5: a-d). Secondary lamella with lamella hyperemia was identified more in fish specimens from Ogbogoro while there was no necrosis identified in Arac specimens.



Fig-5: (a-e) Gills microscopic structures {H and E, x400) (a-b) Indicates Vacillation (VAC) and Necrosis (NEC) (c-d) Hyperemia (Hp) and Haemorrahge (HM) (e) Fusion of secondary lamella (FSL)

DISCUSSION Gill Histopathology

The highest number of histopathological alterations was observed in the gills. These included circulatory disturbances in the form of Hyperamia, haemorrhage, vacuolation and epithelia lifting. Primarily regressive changes in the form of structural alterations and necrosis were also seen. These alterations are not toxicant specific but can be associated with pathogens, and metal pollution in the water (copper)[21].

The gills are sensitive indicators of environmental stress, including exposure to harmful compounds present in aquatic ecosystems as a result of anthropogenic activities [22]. The gills in fish are vulnerable to toxicants and irritants because they are in direct contact with the surrounding water and have a rich blood supply to pick up oxygen for respiration from the water [23]. The gills are probably the most sensitive organs for toxicant exposure as they are actively filter water bound and oxygen.

In the current study histological alterations in varying degrees were identified in gills. These were mostly circulatory disturbances and regressive changes. Circulatory disturbances are related to pathological conditions of blood and tissue fluid flow. Epithelial lifting in focal areas was noted in both fish species. Epithelial lifting is characterized by detachment of epithelial cells due to the outflow of serous fluids into the interstices of gill tissue [18]. This alteration has been observed in various other studies [24, 25, 19, 26].

Epithelial lifting may be a defense mechanism of fish in response to toxicants. The lifting up of epithelium increases the distance through which toxicant has to travel to reach the blood stream [27,15].

Structural alterations in the form of lamella fusion were also identified. Fusion of lamellae is the result of hyperplasia of undifferentiated gill epithelial cells. According to Mallat [14] lamella fusion could be protective in that it diminishes the amount of vulnerable gill surface area. This alteration has previously been identified in fish exposed to pesticides Fish [28, 29, 15], detergents [30] and polluted streams [20].

CONCLUSION

In conclusion, the severity of the alterations in the Gill showed that Ogbogoro commercial fish farm is moderately polluted and not significant (P>0.05) which is considered to belong to the second class of level of alterations as stated by Zimmerli's form of classification.

REFERENCES

1. Dickens, C. W. S., & Graham, P. M. (2002). *African Journal of Aqautic Science*, 1-10

- 2. Olaleye, V. F., & Adewumi, A. A (2001). African Journal of Agricultural Research, 6(6), 1282-1285.
- 3. Skelton, P. (2001). A Complete Guide the *Freshwater Fishes of Southern Africa*. Struik Publishers.
- Anoop, K. R., Sundar, K. S. G., Khan, B. A., & Lal, S. (2009). Common Moorhen *Gallinula chloropus* in the diet of the African catfish *Clarias gariepinus* in Keoladeo Ghana National Park, India. *Indian Birds*, 5(2), 22-23.
- Allison, T. A., & Paul, C. W. (2014). Histological Based Biomonitoring: A Baseline Ecotoxicological Evaluation of New Calabar River Using Chrysichthis Nigrodigitatus. Europe American Journal: International journal of Environment and Pollution Research, 2(3).
- Maltby, L., & Naylor, C. (1990). Preliminary observations on the ecological relevance of the Gammarusscope for growth'assay: effect of zinc on reproduction. *Functional Ecology*, 393-397.
- 7. Crick, F. H., & Orgel, L. E. (1973). Directed Panspermia Icarus, 341-348
- 8. Bradford, A. (2015). Pollution facts and types of pollution
- 9. Skelton, P. (2001). A Complete Guide to the Freshwater Fishes of Southern Africa. South Africa: Struik Publishers.
- 10. Abernathy, C. O., & Donohue, J. M. (1999). Exposure to inorganic arsenic from fish and shellfish.
- Seager, S., Kuchner, M., Hier-Majumder, C. A., & Militzer, B. (2007). Mass-radius relationships for solid exoplanets. *The Astrophysical Journal*, 669(2), 1279.
- Kotsanis, N., & Iliopoulou-Georgudaki, J. (1999) Arsenic induced liver hyperplasia and kidney fibrosis in rainbow trout (Oncorhynchus mykiss) by microinjection technique: A sensitive animal bioassay for environmental metal-toxicity. *Bull Environ Contam Toxicol*, 62, 169-178
- 13. Rainer, F., & Daniel, P. (2014). "*Clarias gariepinus*" in FishBase. March 2014 version.
- 14. Mallatt J. (1985). Fish gill structural changes induced by toxicants and other irritants: a statistical review. *Can. J. Fish. Aquat. Sci, 42, 630-648.*
- 15. Cengiz, E. I. (2006). Gill and kidney histopathology in freshwater fish *Cyprinus carpio* after acute exposure to deltamethrin. *Environ. Toxicol. Pharm.*, 22, 200-204.
- Hinton, B., & Lauren, J. (1990). Integrative Histopathology Approaches to detecting effect of Environmental Stressors on Fishes. *Am. Fish. Soc. Sym.*, 81-51-66
- 17. Akiyoshi, H., & Inoue, A. (2004). Comparative histological study of teleost livers in relation to phylogeny. *Zool.Sci.*, *21*, 841-850.
- Van Dyk, J. C., Marchand, M. J., Pieterse, G. M., Barnhoorn, I. E. J., & Bornman, M. S. (2009). Histological changes in the gills of *Clarias*

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gariepinus from a polluted aquatic system, Gauteng, South Africa. Afr. J. Aquat. Sci., 34(3), 283-291.

- 19. Bernet, D., Schmidt-Posthaus, H., Wahli, T., & Burkhardt-Holm, P. (2004). Evaluation of two monitoring approaches to assess effects of waste water disposal on histological alterations in fish. *Hydrobiologia*, *524*, 53-66.
- Van Dyk, J. C., Marchand, M. J., Pieterse, G. M., Barnhoorn, I. E. J., & Bornman, M. S. (2009). Histological changes in the gills of *Clarias* gariepinus (Teleostei: Clariidae) from a polluted South African urban aquatic system. *Afr. J. Aquat. Sci. 34, 283–291.*
- Skidmore, J. F., & Tovell, P. W. A. (1972). Toxic effects of zinc sulphate on the gills of rainbow trout. *Water Res.*, *6*, 217-230.
- Hinton, D. E., Baumann, P. C., Gardner, G. C., Hawkins, W. E., Hendricks, J. D., Murchelano, R. A., & Okihiro, M. S. (1992). Histopathologic biomarkers. In: Biomarkers: Biochemical, Physiological and Histological Markers of Anthropogenic Stress. Edited by Huggett, R. J., Kimerly, R. A., Mehrle, P. M. (Jr). and Bergman, H.L. Chelsea, M.I., USA: Lewis Publishers
- 23. Roberts, R. J. (2001). *Fish pathology*. W.B. Saunders: Harcourt Publishers Ltd.
- Farsak, B., Yildirir, A., Akyön, Y., Pinar, A., Öç, M., Böke, E., ... & Tokgözoğlu, L. (2000). Detection of Chlamydia pneumoniae andHelicobacter pylori DNA in Human Atherosclerotic Plaques by PCR. *Journal of Clinical Microbiology*, 38(12), 4408-4411.
- 25. Cengiz, E. I., & Unlu, E. (2002). Histopathological changes in the gills of mosquitofish. *Gambusia affinis* exposed to endosulfan. *Bull. Environ. Contam. Toxicol.*, 68 (2), 290-296.
- Vinodhini, R., & Narayanan, M. (2009). Heavy metals induced histopathological alterations in selected organs of *Cyprinus carpio L*. (Common carp). *Int. J. Environ. Res.*, 3(1), 95-100.
- Morgan, M., & Tovell, P. W. A. (1973). The structure of the gill of the trout (*Salmogairdneri*) (Richardson). *Zellforch Mikrosk Anat.*, 142, 147-162
- Jiraungkoorskul, W., Upatham, E. S., Kruatrachue, M., Sahaphong, S., Vichasri-Grams, S., & Pokethitiyook, P. (2002). Histopathological effects of Roundup, a Glyphosate herbicide, on Tilapia (*Oreochromis niloticus*). *ScienceAsia*, 28, 121-127.
- Lara-Ortiz, T., Riveros-Rosas, H., & Aguirre, J. (2003). Reactive oxygen species generated by microbial NADPH oxidase NxA regulate sexual development in *Aspergillus nidulans*. *Mol Microbiol*, 50, 1241-1255.
- Ortiz, J. B., De Canales, M. L. G., & Sarasquete, C. (2003). Histopathological changes induced lindane (gamma-HCH) in various organs of fishes. *Sci. Mar.*, 67 (1), 53-61.